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(21) International Application Number: PCT/US00/14867 (22) International Filing Date: 30 May 2000 (30.05.2000) (30) Priority Data: 09/322,913 01 June 1999 (01.06.1999) US (60) Parent Application or Grant SCHERING CORPORATION [/]; O. DOWLING, Lynette, M. [/]; O. TIMANS, Jacqueline, C. [/]; O. GORMAN, Daniel, M. [/]; O. KASTELEIN, Robert, A. [/]; O. BAZAN, Fernando, J. [/]; O. SCHRAM, David, B. ; O.	Published	
(54) Title: MAMMALIAN RECEPTOR PROTEINS; RELATED REAGENTS AND METHODS (54) Titre: PROTEINES DE RECEPTEUR MAMMALIENNES, REACTIFS ET PROCEDES Y RELATIFS (57) Abstract Nucleic acids encoding mammalian, e.g., primate, receptors, purified receptor proteins and fragments thereof. Antibodies, both polyclonal and monoclonal, are also provided. Methods of using the compositions for both diagnostic and therapeutic utilities are described. (57) Abrégé La présente invention concerne des acides nucléiques codant pour des protéines mammaliennes, par exemple de primates, de récepteurs, de récepteurs purifiées et les fragments de celles-ci. L'invention concerne également des anticorps polyclonaux et monoclonaux. Enfin, l'invention concerne des procédés d'utilisation des compositions à des fins diagnostiques et thérapeutiques.		

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(54) Title: MAMMALIAN RECEPTOR PROTEINS; RELATED REAGENTS AND METHODS

(57) Abstract: Nucleic acids encoding mammalian, e.g., primate, receptors, purified receptor proteins and fragments thereof. Antibodies, both polyclonal and monoclonal, are also provided. Methods of using the compositions for both diagnostic and therapeutic utilities are described.

Description

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MAMMALIAN RECEPTOR PROTEINS; RELATED REAGENTS AND METHODS

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FIELD OF THE INVENTION

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The present invention relates to compositions and methods for affecting mammalian physiology, including immune system function. In particular, it provides methods to regulate development and/or the immune system. Diagnostic and therapeutic uses of these materials are also disclosed.

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BACKGROUND OF THE INVENTION

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Recombinant DNA technology refers generally to techniques of integrating genetic information from a donor source into vectors for subsequent processing, such as through introduction into a host, whereby the transferred genetic information is copied and/or expressed in the new environment. Commonly, the genetic information exists in the form of complementary DNA (cDNA) derived from messenger RNA (mRNA) coding for a desired protein product. The carrier is frequently a plasmid having the capacity to incorporate cDNA for later replication in a host and, in some cases, actually to control expression of the cDNA and thereby direct synthesis of the encoded product in the host. See, e.g., Sambrook, et al. (1989) Molecular Cloning: A Laboratory Manual, (2d ed.) vols. 1-3, CSH Press, NY.

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For some time, it has been known that the mammalian immune response is based on a series of complex cellular interactions, called the "immune network". Recent research has provided new insights into the inner workings of this network. While it remains clear that much of the immune response does, in fact, revolve around the network-like interactions of lymphocytes, macrophages, granulocytes, and other cells, immunologists now generally hold the opinion that soluble proteins, known as lymphokines, cytokines, or monokines, play critical roles in controlling these cellular interactions. Thus, there is considerable interest in the isolation, characterization, and mechanisms of action of cell modulatory factors, an

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5 understanding of which will lead to significant advancements in
the diagnosis and therapy of numerous medical abnormalities,
e.g., immune system disorders.

10 Lymphokines apparently mediate cellular activities in a
5 variety of ways. See, e.g., Paul (ed. 1996) Fundamental
Immunology 3d ed., Raven Press, New York; and Thomson (ed.
1994) The Cytokine Handbook 2d ed., Academic Press, San Diego.
15 They have been shown to support the proliferation, growth,
and/or differentiation of pluripotential hematopoietic stem
20 cells into vast numbers of progenitors comprising diverse
cellular lineages which make up a complex immune system.
Proper and balanced interactions between the cellular
25 components are necessary for a healthy immune response. The
different cellular lineages often respond in a different manner
15 when lymphokines are administered in conjunction with other
agents.

25 Cell lineages especially important to the immune response
include two classes of lymphocytes: B-cells, which can produce
and secrete immunoglobulins (proteins with the capability of
20 recognizing and binding to foreign matter to effect its
removal), and T-cells of various subsets that secrete
30 lymphokines and induce or suppress the B-cells and various
other cells (including other T-cells) making up the immune
network. These lymphocytes interact with many other cell
35 25 types.

40 Research to better understand and treat various immune
disorders has been hampered by the general inability to
maintain cells of the immune system in vitro. Immunologists
45 have discovered that culturing many of these cells can be
30 accomplished through the use of T-cell and other cell
supernatants, which contain various growth factors, including
many of the lymphokines.

50 Various growth and regulatory factors exist which modulate
morphogenetic development. This includes, e.g., the Toll
35 ligands, which signal through binding to receptors which share
structural, and mechanistic, features characteristic of the IL-
1 receptors. See, e.g., Lemaitre, et al. (1996) Cell 86:973-
55 983; and Belvin and Anderson (1996) Ann. Rev. Cell & Devel.

5 Biol. 12:393-416. Other receptors for cytokines are also
known. Often, there are at least two critical subunits in the
functional receptor. See, e.g., Gonda and D'Andrea (1997)
10 Blood 89:355-369; Presky, et al. (1996) Proc. Nat'l Acad. Sci.
5 USA 93:14002-14007; Drachman and Kaushansky (1995) Curr. Opin.
Hematol. 2:22-28; Theze (1994) Eur. Cytokine Netw. 5:353-368;
and Lemmon and Schlessinger (1994) Trends Biochem. Sci. 19:459-
463.

15 From the foregoing, it is evident that the discovery and
10 development of new soluble proteins and their receptors,
including ones similar to lymphokines, should contribute to new
therapies for a wide range of degenerative or abnormal
20 conditions which directly or indirectly involve development,
differentiation, or function, e.g., of the immune system and/or
15 hematopoietic cells. In particular, the discovery and
understanding of novel receptors for lymphokine-like molecules
25 which enhance or potentiate the beneficial activities of other
lymphokines would be highly advantageous. The present
invention provides new receptors for ligands exhibiting
20 similarity to cytokine like compositions and related compounds,
and methods for their use.

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SUMMARY OF THE INVENTION

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The present invention is directed to novel receptors related to cytokine receptors, e.g., primate, cytokine receptor like molecular structures, designated DNAX Cytokine Receptor Subunits (DCRS), and their biological activities. In particular, it provides description of one subunit, designated DCRS2. It includes nucleic acids coding for the polypeptides themselves and methods for their production and use. The nucleic acids of the invention are characterized, in part, by their homology to cloned complementary DNA (cDNA) sequences enclosed herein.

The present invention provides a composition of matter selected from: a substantially pure or recombinant DCRS2 polypeptide comprising at least three distinct nonoverlapping segments of at least four amino acids identical to segments of SEQ ID NO: 2; a substantially pure or recombinant DCRS2 polypeptide comprising at least two distinct nonoverlapping segments of at least five amino acids identical to segments of SEQ ID NO: 2; a natural sequence DCRS2 comprising mature SEQ ID NO: 2; or a fusion polypeptide comprising DCRS2 sequence. In certain embodiment, the invention embraces such a substantially pure or isolated antigenic DCRS2 polypeptide, wherein the distinct nonoverlapping segments of identity: include one of at least eight amino acids; include one of at least four amino acids and a second of at least five amino acids; include at least three segments of at least four, five, and six amino acids, or include one of at least twelve amino acids. Other embodiments include wherein the: DCRS2 polypeptide: comprises a mature sequence of Table 1; is an unglycosylated form of DCRS2; is from a primate, such as a human; comprises at least seventeen amino acids of SEQ ID NO: 2; exhibits at least four nonoverlapping segments of at least seven amino acids of SEQ ID NO: 2; is a natural allelic variant of DCRS2; has a length at least about 30 amino acids; exhibits at least two non-overlapping epitopes which are specific for a primate DCRS2; is glycosylated; has a molecular weight of at least 30 kD with natural glycosylation; is a synthetic polypeptide; is attached to a solid substrate; is conjugated to another chemical moiety;

5 is a 5-fold or less substitution from natural sequence; or is a
deletion or insertion variant from a natural sequence. Still
other embodiments include a composition comprising: a
10 substantially pure DCRS2 and another Interferon Receptor family
5 member; a sterile DCRS2 polypeptide; the DCRS2 polypeptide and
a carrier, wherein the carrier is: an aqueous compound,
including water, saline, and/or buffer; and/or formulated for
oral, rectal, nasal, topical, or parenteral administration.
15 Fusion polypeptide embodiments include those comprising: mature
10 protein sequence of Table 1; a detection or purification tag,
including a FLAG, His6, or Ig sequence; or sequence of another
interferon receptor protein. Kit embodiments include those
20 comprising such a polypeptide, and: a compartment comprising
the protein or polypeptide; or instructions for use or disposal
15 of reagents in the kit.

25 Binding compound embodiments include, e.g., a binding
compound comprising an antigen binding site from an antibody,
which specifically binds to a natural DCRS2 polypeptide,
wherein: the binding compound is in a container; the DCRS2
20 polypeptide is from a human; the binding compound is an Fv,
Fab, or Fab2 fragment; the binding compound is conjugated to
another chemical moiety; or the antibody: is raised against a
peptide sequence of a mature polypeptide of Table 1; is raised
against a mature DCRS2; is raised to a purified human DCRS2; is
35 25 immunoselected; is a polyclonal antibody; binds to a denatured
DCRS2; exhibits a K_d to antigen of at least 30 μM ; is attached
to a solid substrate, including a bead or plastic membrane; is
in a sterile composition; or is detectably labeled, including a
40 radioactive or fluorescent label. Kits include those
30 comprising the binding compound, and: a compartment comprising
the binding compound; or instructions for use or disposal of
reagents in the kit.

45 Methods are provided, e.g., of producing an
antigen:antibody complex, comprising contacting under
35 appropriate conditions a primate DCRS2 polypeptide with a
described antibody, thereby allowing the complex to form. This
includes wherein: the complex is purified from other interferon
50 receptors; the complex is purified from other antibody; the

5 contacting is with a sample comprising an interferon; the
contacting allows quantitative detection of the antigen; the
contacting is with a sample comprising the antibody; or the
contacting allows quantitative detection of the antibody.

10 5 Various related compositions are provided, e.g., a
composition comprising: a sterile binding compound, as
described, or the described binding compound and a carrier,
wherein the carrier is: an aqueous compound, including water,
15 saline, and/or buffer; and/or formulated for oral, rectal,
20 nasal, topical, or parenteral administration.

Nucleic acid embodiments include, e.g., an isolated or
recombinant nucleic acid encoding the DCRS2 polypeptide,
wherein the: DCRS2 is from a human; or the nucleic acid:
encodes an antigenic peptide sequence of Table 1; encodes a
15 plurality of antigenic peptide sequences of Table 1; exhibits
identity over at least thirteen nucleotides to a natural cDNA
25 encoding the segment; is an expression vector; further
comprises an origin of replication; is from a natural source;
comprises a detectable label; comprises synthetic nucleotide
20 sequence; is less than 6 kb, preferably less than 3 kb; is from
a primate; comprises a natural full length coding sequence; is
30 a hybridization probe for a gene encoding the DCRS2; or is a
PCR primer, PCR product, or mutagenesis primer. Other
embodiments of the invention include a cell or tissue
35 25 comprising the described recombinant nucleic acid. Preferably,
the cell is: a prokaryotic cell; a eukaryotic cell; a bacterial
cell; a yeast cell; an insect cell; a mammalian cell; a mouse
cell; a primate cell; or a human cell.

40 Kit embodiments include those comprising a described
30 nucleic acid, and: a compartment comprising the nucleic acid; a
compartment further comprising a primate DCRS2 polypeptide; or
instructions for use or disposal of reagents in the kit.

45 Alternative nucleic acid embodiments include a nucleic
acid which: hybridizes under wash conditions of 30 minutes at
35 30° C and less than 2M salt to the coding portion of SEQ ID NO:
1; or exhibits identity over a stretch of at least about 30
nucleotides to a primate DCRS2. Preferred embodiments include
50 those wherein: the wash conditions are at 45° C and/or 500 mM

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salt; the wash conditions are at 55° C and/or 150 mM salt; the stretch is at least 55 nucleotides; or the stretch is at least 75 nucleotides.

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5 Other methods include those of modulating physiology or development of a cell or tissue culture cells comprising contacting the cell with an agonist or antagonist of a mammalian DCRS2. Preferably, the cell is transformed with a nucleic acid encoding a DCRS2 and another cytokine receptor subunit.

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DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

OUTLINE

- I. General
- 5 II. Activities
- 10 III. Nucleic acids
 - A. encoding fragments, sequence, probes
 - B. mutations, chimeras, fusions
 - C. making nucleic acids
 - 10 D. vectors, cells comprising
- 15 IV. Proteins, Peptides
 - A. fragments, sequence, immunogens, antigens
 - B. muteins
 - C. agonists/antagonists, functional equivalents
 - 15 D. making proteins
- 20 V. Making nucleic acids, proteins
 - A. synthetic
 - B. recombinant
 - C. natural sources
- 20 VI. Antibodies
 - A. polyclonals
 - B. monoclonal
 - 25 C. fragments; Kd
 - D. anti-idiotypic antibodies
 - 25 E. hybridoma cell lines
- VII. Kits and Methods to quantify DCRS2
 - A. ELISA
 - B. assay mRNA encoding
 - 30 C. qualitative/quantitative
 - 30 D. kits
- VIII. Therapeutic compositions, methods
 - A. combination compositions
 - B. unit dose
 - C. administration
- 35 IX. Screening
- X. Ligands

I. General

40 The present invention provides the amino acid sequence and DNA sequence of mammalian, herein primate, cytokine receptor-like subunit molecules, this one designated DNAX Cytokine Receptor Subunit 2 (DCRS2) having particular defined properties, both structural and biological. Various cDNAs encoding these molecules were obtained from primate, e.g., 45 human, cDNA sequence libraries. Other primate or other mammalian counterparts would also be desired.

50 Some of the standard methods applicable are described or referenced, e.g., in Maniatis, et al. (1982) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring

5 Harbor Press; Sambrook, et al. (1989) Molecular Cloning: A
Laboratory Manual, (2d ed.), vols. 1-3, CSH Press, NY; Ausubel,
et al., Biology, Greene Publishing Associates, Brooklyn, NY; or
10 Ausubel, et al. (1987 and periodic supplements) Current
5 Protocols in Molecular Biology, Greene/Wiley, New York; each of
which is incorporated herein by reference.

Nucleotide (SEQ ID NO: 1) and corresponding amino acid
sequence (SEQ ID NO: 2) of a human DCRS2 coding segment is
15 shown in Table 1. It is likely that there is at least one
10 splice variant with a longer intracellular domain, and will
probably exhibit characteristic signaling motifs. The
predicted signal sequence is indicated, but may depend on cell
20 type, or may be a few residues in either direction. Potential
N glycosylation sites are at Asparagine residues 6, 24, 58,
15 118, 157, 209, and 250. Disulfide linkages are likely to be
found between cysteine residues at positions 29 and 78; and a
conserved C_CXW motif is found at positions 110/121/123. The
25 tryptophan at 219; and the WxxWS motif from 281-285 are
notable. The segment from about 1-101 is an Ig domain; from
20 about 102-195 is a cytokine binding domain 1; from about 196-
30 297 is a cytokine binding domain 2; from about 298-330 is a
linker; from about 331-353 is a transmembrane segment; and from
about 354-361 is an intracellular domain. These sites and
boundaries are notable.

35 25 The reverse translation nucleic acid sequence is provided
in Table 2.

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Table 1: Nucleotide and polypeptide sequences of DNAX Cytokine Receptor Subunit like embodiments (DCRS2). Primate, e.g., human embodiment (see SEQ ID NO: 1 and 2). Predicted signal sequence indicated, but may vary by a few positions and depending upon cell type.

5	5	atg aat cag gtc act att caa tgg gat gca gta ata gcc ctt tac ata	48
10	10	Met Asn Gln Val Thr Ile Gln Trp Asp Ala Val Ile Ala Leu Tyr Ile	
		-20	-15
		-10	
10	10	ctc ttc agc tgg tgt cat gga gga att aca aat ata aac tgc tct ggc	96
		Leu Phe Ser Trp Cys His Gly Gly Ile Thr Asn Ile Asn Cys Ser Gly	
		-5	-1
		1	5
15	15	cac atc tgg gta gaa cca gcc aca att ttt aag atg ggt atg aat atc	144
		His Ile Trp Val Glu Pro Ala Thr Ile Phe Lys Met Gly Met Asn Ile	
		10	15
		20	25
20	20	tct ata tat tgc caa gca gca att aag aac tgc caa cca agg aaa ctt	192
		Ser Ile Tyr Cys Gln Ala Ala Ile Lys Asn Cys Gln Pro Arg Lys Leu	
		30	35
		40	
25	25	cat ttt tat aaa aat ggc atc aaa gaa aga ttt caa atc aca agg att	240
		His Phe Tyr Lys Asn Gly Ile Lys Glu Arg Phe Gln Ile Thr Arg Ile	
		45	50
		55	
25	25	aat aaa aca aca gct cgg ctt tgg tat aaa aac ttt ctg gaa cca cat	288
		Asn Lys Thr Thr Ala Arg Leu Trp Tyr Lys Asn Phe Leu Glu Pro His	
		60	65
		70	
25	30	gct tct atg tac tgc act gct gaa tgt ccc aaa cat ttt caa gag aca	336
		Ala Ser Met Tyr Cys Thr Ala Glu Cys Pro Lys His Phe Gln Glu Thr	
		75	80
		85	
30	35	ctg ata tgt gga aaa gac att tct tct gga tat ccg cca gat att cct	384
		Leu Ile Cys Gly Lys Asp Ile Ser Ser Gly Tyr Pro Pro Asp Ile Pro	
		90	95
		100	105
30	40	gat gaa gta acc tgt gtc att tat gaa tat tca ggc aac atg act tgc	432
		Asp Glu Val Thr Cys Val Ile Tyr Glu Tyr Ser Gly Asn Met Thr Cys	
		110	115
		120	
35	45	acc tgg aat gct ggg aag ctc acc tac ata gac aca aaa tac gtg gta	480
		Thr Trp Asn Ala Gly Lys Leu Thr Tyr Ile Asp Thr Lys Tyr Val Val	
		125	130
		135	
35	45	cat gtg aag agt tta gag aca gaa gaa gag caa cag tat ctc acc tca	528
		His Val Lys Ser Leu Glu Thr Glu Glu Glu Gln Gln Tyr Leu Thr Ser	
		140	145
		150	
40	50	agc tat att aac atc tcc act gat tca tta caa ggc ggc aag aag tac	576
		Ser Tyr Ile Asn Ile Ser Thr Asp Ser Leu Gln Gly Gly Lys Lys Tyr	
		155	160
		165	
40	55	ttg gtt tgg gtc caa gca gca aac gca cta ggc atg gaa gag tca aaa	624
		Leu Val Trp Val Gln Ala Ala Asn Ala Leu Gly Met Glu Glu Ser Lys	
		170	175
		180	185

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caa ctg caa att cac ctg gat gat ata gtg ata cct tct gca gcc gtc 672
 Gln Leu Gln Ile His Leu Asp Asp Ile Val Ile Pro Ser Ala Ala Val
 190 195 200

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att tcc agg gct gag act ata aat gct aca gtg ccc aag acc ata att 720
 Ile Ser Arg Ala Glu Thr Ile Asn Ala Thr Val Pro Lys Thr Ile Ile
 205 210 215

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tat tgg gat agt caa aca aca att gaa aag gtt tcc tgt gaa atg aga 768
 Tyr Trp Asp Ser Gln Thr Thr Ile Glu Lys Val Ser Cys Glu Met Arg
 220 225 230

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tac aag gct aca aca aac caa act tgg aat gtt aaa gaa ttt gac acc 816
 Tyr Lys Ala Thr Thr Asn Gln Thr Trp Asn Val Lys Glu Phe Asp Thr
 235 240 245

aat ttt aca tat gtg caa cag tca gaa ttc tac ttg gag cca aac att 864
 Asn Phe Thr Tyr Val Gln Gln Ser Glu Phe Tyr Leu Glu Pro Asn Ile
 250 255 260 265

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aag tac gta ttt caa gtg aga tgt caa gaa aca ggc aaa agg tac tgg 912
 Lys Tyr Val Phe Gln Val Arg Cys Gln Glu Thr Gly Lys Arg Tyr Trp
 270 275 280

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cag cct tgg agt tca ccg ttt ttt cat aaa aca cct gaa aca gtt ccc 960
 Gln Pro Trp Ser Ser Pro Phe Phe His Lys Thr Pro Glu Thr Val Pro
 285 290 295

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cag gtc aca tca aaa gca ttc caa cat gac aca tgg aat tct ggg cta 1008
 Gln Val Thr Ser Lys Ala Phe Gln His Asp Thr Trp Asn Ser Gly Leu
 300 305 310

aca gtt gct tcc atc tct aca ggg cac ctt act tct gac aac aga gga 1056
 Thr Val Ala Ser Ile Ser Thr Gly His Leu Thr Ser Asp Asn Arg Gly
 315 320 325

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gac att gga ctt tta ttg gga atg atc gtc ttt gct gtt atg ttg tca 1104
 Asp Ile Gly Leu Leu Leu Gly Met Ile Val Phe Ala Val Met Leu Ser
 330 335 340 345

att ctt tct ttg att ggg ata ttt aac aga tca ttc ccg aac tgg gat 1152
 Ile Leu Ser Leu Ile Gly Ile Phe Asn Arg Ser Phe Pro Asn Trp Asp
 350 355 360

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taa 1155

50 MNQVTIQWDAVIALYILFSWCHGGITNINCSGHIWVEPATIFKMGMNISYCOAAIKNCQPRKLFYKNGIKERF
 QITRINKTARLWYKNFLEPHASMYCTAECPKHFQETLICGDISSGYPPDIPDEVTCVIYEYSGNMTCTWNAGK
 40 LTYIDTKYVVHVKSLETEEEQQYLTSSYINISTDSLQGGKKYLWVWQAANALGMEESKQLQIHLDDIVIPSAAVI
 SRAETINATVPKTIYWDSQTTIEKVSCEMRYKATTNQTWNVKEFDTNFTYVQOSEFYLEPNIKYVPQVRCQETG
 KRYWQPWSSPPFHKTPEVTPQVTSKAFQHDWNSGLTVASISTGHLTSDNRGDIIGLLGMIVFAVMLSILSLIGI
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5 Table 2: Reverse Translation of primate, e.g., human, DCRS2 (SEQ ID NO: 3):

	atgaaycarg tnacnathca rtgggaygcn gtnathgcn y tntayathyt nttywsntgg 60
5	tgycayggng gnathacnaa yathaaytgy wsnngncaya thtgggtnga rccngcnacn 120
10	athttyaara tgggnatgaa yathwsnath taytgycarg cngcnathaa raaytgycar 180
10	ccnmnaary tncayttyta yaaraayggn athaargarm gnttycarat hacnmgnath 240
15	aayaaracna cngcnmgnyt ntgggtayaar aayttytng arccncaygc nwsnatgtay 300
15	tgyacngcng artgyccnaa rcayttycar garacnytna thtgyggnaa rgayathwsn 360
15	wsnngntayc cncngayat hccngaygar gtnacntgyg tnathtayga rtaywsnggn 420
20	aayatgacnt gyacntggaa ygcnggnaar ytnacntaya thgayacnaa rtaygtngtn 480
20	caygtnaarw snytngarac ngargargar carcarty tnacwnswn ntayathaay 540
20	athwsnacng aywsnytnca rggnggnaar aartaytng tntgggtnga rgcngcnaay 600
25	gcnytnggna tggargarws naarcarytn carathcayy tngaygayat hgtathccn 660
25	wsnngcngcng tnatwsnmg ngcngaracn athaaygna cngtncnaa racnathath 720
25	taytgggayw snacaracnac nathgaraar gtnwsntgyg aratgmgnta yaargcnacn 780
25	acnaaycara cntggaaygt naargartty gayacnaayt tyacntaygt ncarcarwsn 840
30	garttytayy tngarccnaa yathaartay gtnttycarg tnmngtgyca rgaracnggn 900
30	aarmgntayt ggcarrcentg gwsnwsncn ttytycaya aracncnga racngtncn 960
35	cargtnacnw snaargcntt ycarcaygay acntggaayw snggnytnac ngtngcnwsn 1020
30	athwsnacng gncayytnac nwsngayaay mngngngaya thggnytnyt nytnggnatg 1080
40	athgtnttyg cngtntgyt nwsnathytn wsnynathg gnathttyaa ymgnwsntty 1140
	ccnaaytggg ay 1152

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Table 3: Alignment of various cytokine receptor subunits. Human NR6 sequence (hNR6) is SEQ ID NO: 4 (see Elson, et al. (1998) J. Immunol. 161:1371-1379; GenBank Accession number AF059293; also described by Douglas J. Hilton (Australia)); mouse NR6 sequence (mNR6) is SEQ ID NO: 5. Human p40 (hp40) is SEQ ID NO: 6 (see GenBank M65272); mouse p40 is SEQ ID NO: 7 (see GenBank S82421). Mouse Ebi3 (mEbi3) is SEQ ID NO: 8 (see GenBank AF013114); human Ebi3 (hEbi3) is SEQ ID NO: 9 (see GenBank L08187). Mouse IL-11 Receptor subunit alpha (mIL-11Ra) is SEQ ID NO: 10 (see GenBank U14412); human IL-11 Receptor subunit alpha (hIL-11Ra) is SEQ ID NO: 11 (see GenBank U32324). Human IL-6 Receptor subunit alpha (hIL-6Ra) is SEQ ID NO: 12 (see GenBank X58298); mouse IL-6 Receptor subunit alpha (mIL-6Ra) is SEQ ID NO: 13 (see GenBank X51975).

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hNR6      MPACRRGPAQAQSARRPPFLLPLLLLLCVLGAPRAGSGAHTAVISPODPTL
mNR6      -----RPLSSLWSPLLLCVLGVPRGGGAHTAVISPODPTL
hp40      -----MCHQQLVISWPSLVFLASPLVAIWELKKDVYVVELDWYP
mp40      -----MCPQKLTISWFAIVLLVSPLMAMWELEKDVYVVEVDWTP
mEbi3     -----
hEbi3     -----
mIL-11Ra  -----MSSSCSGLTRVLVAVATALVSSSPCPQAWGPPGVQYG
hIL-11Ra  -----MSSSCSGLSRVLVAVATALVSASSPCQAWGPPGVQYG
hIL-6Ra   -----MLAVGCALLAALLAAPGAALAPRR--CPAQEVARGVLT
mIL-6Ra   -----MLTVGCTLLVALLAAPAVALVLGS--CRALEVANGTVTS
hAS11     -----MNQVTIQWDAVIALYILFSWCHGGITNINCSGHIWVEPATIFK

hNR6      -LIGSSLLATCSVHGDPGATAEGLYWTLNRRLLPPELSRVLNASTLALA
mNR6      -LIGSSLQATCSIHGDTGATAEGLYWTLNRRLLPPELSRVLNASTLALA
hp40      DAPGEMVVLTCDTPEED-----GITWTLD-----QSSEVLGSGKTLT
mp40      DAPGETVNLTCDTPEED-----DITWTSD-----QRHGVIGSGKTLT
mEbi3     -----MSKLLF
hEbi3     -----MTPQLL
mIL-11Ra  -QPGRPVMLCCPGVSAG----TPVSWFRDGS-R-LLQGPDSGLGHLV
hIL-11Ra  -QPGRSVKLCPCGVTAG----DPVSWFRDGEK-K-LLQGPDSGLGHELV
hIL-6Ra   -LPGDSVTLTCPGVEPED---NATVHWVLRKPAAG-SHPSRWAGMGRRL
mIL-6Ra   -LPGATVTLICPGKEAAG---NVTIHWVYS---G-SQNREWTGNTLV
hAS11     --MGMNISYQAAIKNCQ--PRKLHPYKNGIKER-FQITRINKTTARLW

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hNR6 -PEKPVNISCSWKNMKD-LTCRWTPGAHGETFL--HTNYSLKYLKRWYG-
mNR6 -PEKPFNISCSWRNMKD-LTCRWTPGAHGETFL--HTNYSLKYLKRWYG-
hp40 -EPKNTFLRCEAKNYSGRFTCWLLTTISTDLTFSVKSSRGSSDPQGVTCG
5 mp40 -FKNKTFLKCEAPNYSGRFTCSWLVRQNMMDLKFNIKSSSSSPDSRAVTCG
mEbi3 -----PRVQCHASRYPAVDCSWTFLQAPNSTR--STSFIAIYRLGVATQ
10 hEbi3 -----PRVQCRASRYPIAVDCSWTLPAPNSTR--PVSFIAIYRLGMAAR
mIL-11R -PPARPEVSCQAVDYEN-FSCTWSPGQVSGLPTRYLTSYRKKTLPGAESQ
hIL-11R -PPARPVVSCQAADYEN-FSCTWSPSQISGLPTRYLTSYRKKTVLGADSQ
10 hIL-6Ra -PPEEPQLSCFRKSPLSNVVCEWGRSTPSLTT---KAVLLVRKQPNSP-
mIL-6Ra -PPEEPKLSCFRKNPLVNAICEWRPSSTPSPTT---KAVLFAKKINTNG
hAS11 -PDIPDEVTCVIEYSGNMTCTWAGKLTYYIDT---KYVVHVKSLETE-

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15 hNR6 QDN-----TCEEYHTVGPHSCHIPKDLALF-TPYEIWEATNRLGSA-
mNR6 QDN-----TCEEYHTVGPHSCHIPKDLALF-TPYEIWEATNRLGSA-
hp40 AATLSAERVRGDNKEYE-YSEVCEQEDSACPAEESLPIEVMVDVAVHKLKY
20 mp40 MASLSAEKVTLDRDYEKYSVSCQEDVTCPTAEETLPIELALEARQONKY
mEbi3 QQS-----QPCLQRSQP-ASRCTIPDVHVFSTVPMYMLNVTAVHPGGA--
20 hEbi3 GHS-----WPCLQQTPT-STSCITIDVQLFSMAPYVNLVTAVHPWGS--
mIL-11Ra RESP-STGPWPCCQDPLE-ASRCVVHGAEFWS--EYRINVTEVNSLGA--
hIL-11Ra RRSR-STGPWPCCQDPLG-AARCVVHGAEFWS--QYRINVTEVNLGA--
hIL-6Ra AED---FQEPCCQYSQESQKFSQCLAVPEGDS-SFYIVSMCVASSVGSK-
mIL-6Ra KSD---FQVPCQYSQQLKSFSCQVEILEGDK-VYHIVSLCVANSVGSK-
25 hAS11 -----EEQYLTSSYINISTDSLQGGK--KYLVVWQAANALGME-

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hNR6 RSDVLTLDILDVVTTDPPPDVHVS RVGGLEDQLSVRWVSPPALK--DFLF
mNR6 RSDVLTLDVLDVTTDPPPDVHVS RVGGLEDQLSVRWVSPPALK--DFLF
30 hp40 ENYTSFFIRDIIKPDPPKNLQKPLKNSR-QVEVSWEYPTWSTPHSYF
mp40 ENYTSFFIRDIIKPDPPKNLQMKPLKNS--QVEVSWEYSDSWSTPHSYF
30 mEbi3 SSSLLAFVAERIIKPDPEGVRLRTAGQR---LQVLWHPPASWPF-PDIF
hEbi3 SSSFVFFITEHIIKPDPEGVRLSPLAERH--VQVQWEPGGSWPF-PRIF
mIL-11Ra STCLLDVRLQSILRPDPQGLRVESVPGYPRRLHASWTYPASWRR-QPHF
35 hIL-11Ra STRLLDVSLQSILRPDPQGLRVESVPGYPRRLRASWTYPASWPC-QPHF
hIL-6Ra FSKTQTFQCGGILQDPPANITVTAVARNRRLSVTWQDPSWN--SSFY
35 mIL-6Ra SSHNEAFHSLKMVQDPPANLVSAIPGRPRWLKVSWOHPETWD--PSYY
hAS11 ESKQLQIHLDDVIPSAAVISRAETINATVPKTI IYWDSQTTIE-----

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40 hNR6 QAKYQIRYRVEDSVDWKVV---DDVSNQTSCLRLAGLKPQ-TVYFVQVRCN
mNR6 QAKYQIRYRVEDSVDWKVV---DDVSNQTSCLRLAGLKPQ-TVYFVQVRCN
40 hp40 SLTFCVQVQGKSK--RE-----KKDRVFTDKTSATVICRKNASISVRAQ
mp40 SLKFFVRIQRKKEKMKETEBCGNQKGAFLVEKTS TEVQCK--GGNVCVQAQ
45 mEbi3 SLKYRLRYRRRGASHFR-----QVGPTEATFTLRSKPHAKYCIQVSAQ
hEbi3 SLKYWIRYKRQGAARFH-----RVGPIEATSFILRAVRPRARYVQVAAQ
mIL-11Ra LLKFRLQYRPAQHPAWS-----TVEPIGLEEVITDVTAG-LPHAVRVSAR
hIL-11R LLKFRLQYRPAQHPAWS-----TVEPAGLEEVITDAVAG-LPHAVRVSAR
45 hIL6-Ra RLRFELRYRAERSKTPFTW---MVKDLQHHCVIHDASG-LRHVVQLRAQ
mIL6-Ra LLQFQLRYRFPVWSKEFTVL---LLPVAQYQCVIHDALRG-VKHVVQVRGK
50 hAS11 KVSCEMRYKATTNQTWNVK--EFDTNFTYVQSEFYLEPNIKYVQVRCQ

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hNR6 PFGIYGSKKAGIWEWSHPTAASTPRSE-RPGPGGGACE--PRGGEPSGG
mNR6 PFGIYGSKKAGIWEWSHPTAASTPRSE-RPGPGGGVCE--PRGGEPSGG
hp40 DRYYSSS-----WSEWASVPCS-----
mp40 DRYYNSS-----CSKWACVPCRVR-----
mEbi3 DLTDYGK-----PSDWSLPGQVESAPHKP-----
hEbi3 DLTDYGE-----LSDWSLPATATMSLGK-----
mIL-11Ra DFLDAGT-----WSAWSPEAWGTPSTGLLQDEIPDWSQGHGQQLAVVAQ
hIL-11Ra DFLDAGT-----WSTWSPEAWGTPSTGTIPKEIPAWGQLHTQP--EVEPQ
hIL-6Ra EEFQGE-----WSEWSPEAMGTPWTES-RSPPAENEVS-TPMQALTTNK
mIL-6Ra EELDLGQ-----WSEWSPEVTGTPWIAEPRITTPAGILWNPTQVSVEDSAN
hAS11 ETGKRY-----WQPWSSPFHKTPTVPOVTSKAFQHD-----TWNSS

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hNR6 PVREELKQFLGWLKKHAYCSNLSFRLYDQWRAMQKSHKTRNQ--VLPD
mNR6 PVREELKQFLGWLKKHAYCSNLSFRLYDQWRAMQKSHKTRNQDEGILPS
hp40 -----
mp40 -----
mEbi3 -----
hEbi3 -----
mIL-11Ra EDSLAPARPSLQPPRPLDHRDPLEQVAVLASLGFSCGLAVGALALGL
hIL-11Ra VDSPAPRPSLQPHPRLLDHRDSVEQVAVLASLGLSFLGLVAGALALGL
hIL-6Ra DDDNILFRDSANATSLPVQDSSSVPLPTFLVAGGSLAFGLLCAIVLRF
mIL-6Ra HEDQYESSTEATSVLAPVQESSMSLPTFLVAGGSLAFGLLVCVFIILRL
hAS11 LTVASISTGHLSDNR-GDIGLLGMIVFAVMLSILSLIGIFN--RSPFN

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hNR6 KL-----
mNR6 GRRGAARGPAG-----
hp40 -----
mp40 -----
mEbi3 -----
hEbi3 -----
mIL-11Ra WLRLRRSGKEG-----PQKPGLLAPMIP-----
hIL-11Ra WLRLRRGGKDG-----SPKPGFLASVIP-----
hIL-6Ra KKTWKLRLALKEGKTSMHPP--YSLGQLVPERPRPTVPLVPLISPPVSPSS
mIL-6Ra KQKWSEAEKESKTTSPPPPPYSLGPLK-----TFLVPLLTPHSS--
hAS11 WD-----

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hNR6 -----
mNR6 -----
hp40 -----
mp40 -----
mEbi3 -----
hEbi3 -----
mIL-11Ra -----VEKLPGIPNLQRTPENFS--
hIL-11Ra -----VDRRPGAPNL-----
hIL-6Ra LGSNTSSHNRPDARDPRSPYDISNTDYFFPR
mIL-6Ra -GSDNTVNHSCLGVRDAQSPYDNSNRDYLFFPR
hAS11 -----

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Table 3 shows comparison of the available sequences of primate and rodent receptors with the primate, e.g., human DCRS2 (AS11). The DCRS2 shows particular similarity to the IL-11 receptor subunit alpha, though it may be aligned with the p40 and IL-6 receptor alpha subunits. It is likely an alpha subunit, and thus should bind to ligand without the need for a beta subunit. The biology is likely to be similar to the IL-6 receptor subunit.

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As used herein, the term DCRS2 shall be used to describe a protein comprising the amino acid sequence shown in Table 1. In many cases, a substantial fragment thereof will be functionally or structurally equivalent, including, e.g., an extracellular or intracellular domain. The invention also includes a protein variation of the respective DCRS2 allele whose sequence is provided, e.g., a mutein or soluble extracellular construct. Typically, such agonists or antagonists will exhibit less than about 10% sequence differences, and thus will often have between 1- and 11-fold substitutions, e.g., 2-, 3-, 5-, 7-fold, and others. It also encompasses allelic and other variants, e.g., natural polymorphic, of the protein described. Typically, it will bind to its corresponding biological ligand, perhaps in a dimerized state with an alpha receptor subunit, with high affinity, e.g., at least about 100 nM, usually better than about 30 nM, preferably better than about 10 nM, and more preferably at better than about 3 nM. The term shall also be used herein to refer to related naturally occurring forms, e.g., alleles, polymorphic variants, and metabolic variants of the mammalian protein. Preferred forms of the receptor complexes will bind the appropriate ligand with an affinity and selectivity appropriate for a ligand-receptor interaction.

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This invention also encompasses combinations of proteins or peptides having substantial amino acid sequence identity with the amino acid sequence in Table 1. It will include sequence variants with relatively few substitutions, e.g., preferably less than about 3-5.

A substantial polypeptide "fragment", or "segment", is a stretch of amino acid residues of at least about 8 amino acids,

5 amino acids, often at least 14 amino acids, more often at least
16 amino acids, typically at least 18 amino acids, more
typically at least 20 amino acids, usually at least 22 amino
10 acids, more usually at least 24 amino acids, preferably at
5 least 26 amino acids, more preferably at least 28 amino acids,
and, in particularly preferred embodiments, at least about 30
or more amino acids. Sequences of segments of different
15 proteins can be compared to one another over appropriate length
stretches. In many situations, fragments may exhibit
20 functional properties of the intact subunits, e.g., the
extracellular domain of the transmembrane receptor may retain
the ligand binding features, and may be used to prepare a
soluble receptor-like complex.

Amino acid sequence homology, or sequence identity, is
15 determined by optimizing residue matches. In some comparisons,
gaps may be introduced, as required. See, e.g., Needleham, et
25 al., (1970) J. Mol. Biol. 48:443-453; Sankoff, et al., (1983)
chapter one in Time Warps, String Edits, and Macromolecules:
The Theory and Practice of Sequence Comparison, Addison-Wesley,
20 Reading, MA; and software packages from IntelliGenetics,
Mountain View, CA; and the University of Wisconsin Genetics
30 Computer Group (GCG), Madison, WI; each of which is
incorporated herein by reference. This changes when
considering conservative substitutions as matches.
35 Conservative substitutions typically include substitutions
within the following groups: glycine, alanine; valine,
isoleucine, leucine; aspartic acid, glutamic acid; asparagine,
glutamine; serine, threonine; lysine, arginine; and
40 phenylalanine, tyrosine. Homologous amino acid sequences are
30 intended to include natural allelic and interspecies variations
in the cytokine sequence. Typical homologous proteins or
peptides will have from 50-100% homology (if gaps can be
45 introduced), to 60-100% homology (if conservative substitutions
are included) with an amino acid sequence segment of Table 1.
35 Homology measures will be at least about 70%, generally at
least 76%, more generally at least 81%, often at least 85%,
50 more often at least 88%, typically at least 90%, more typically
at least 92%, usually at least 94%, more usually at least 95%,

5 preferably at least 96%, and more preferably at least 97%, and
in particularly preferred embodiments, at least 98% or more.
The degree of homology will vary with the length of the
10 compared segments. Homologous proteins or peptides, such as
5 the allelic variants, will share most biological activities
with the embodiments described in Table 1.

As used herein, the term "biological activity" is used to
describe, without limitation, effects on inflammatory
15 responses, innate immunity, and/or morphogenic development by
10 cytokine-like ligands. For example, these receptors should
mediate phosphatase or phosphorylase activities, which
activities are easily measured by standard procedures. See,
20 e.g., Hardie, et al. (eds. 1995) The Protein Kinase FactBook
vols. I and II, Academic Press, San Diego, CA; Hanks, et al.
15 (1991) Meth. Enzymol. 200:38-62; Hunter, et al. (1992) Cell
70:375-388; Lewin (1990) Cell 61:743-752; Pines, et al. (1991)
25 Cold Spring Harbor Symp. Quant. Biol. 56:449-463; and Parker,
et al. (1993) Nature 363:736-738. The receptors, or portions
thereof, may be useful as phosphate labeling enzymes to label
20 general or specific substrates. The subunits may also be
functional immunogens to elicit recognizing antibodies, or
30 antigens capable of binding antibodies.

The terms ligand, agonist, antagonist, and analog of,
e.g., a DCRS2, include molecules that modulate the
35 characteristic cellular responses to cytokine ligand proteins,
as well as molecules possessing the more standard structural
binding competition features of ligand-receptor interactions,
e.g., where the receptor is a natural receptor or an antibody.
40 The cellular responses likely are typically mediated through
30 receptor tyrosine kinase pathways.

Also, a ligand is a molecule which serves either as a
natural ligand to which said receptor, or an analog thereof,
45 binds, or a molecule which is a functional analog of the
natural ligand. The functional analog may be a ligand with
35 structural modifications, or may be a wholly unrelated molecule
which has a molecular shape which interacts with the
appropriate ligand binding determinants. The ligands may serve
50 as agonists or antagonists, see, e.g., Goodman, et al. (eds.

5 1990) Goodman & Gilman's: The Pharmacological Bases of Therapeutics, Pergamon Press, New York.

10 Rational drug design may also be based upon structural studies of the molecular shapes of a receptor or antibody and other effectors or ligands. See, e.g., Herz, et al. (1997) J. Recept. Signal Transduct. Res. 17:671-776; and Chaiken, et al. (1996) Trends Biotechnol. 14:369-375. Effectors may be other proteins which mediate other functions in response to ligand binding, or other proteins which normally interact with the receptor. One means for determining which sites interact with specific other proteins is a physical structure determination, e.g., x-ray crystallography or 2 dimensional NMR techniques. These will provide guidance as to which amino acid residues form molecular contact regions. For a detailed description of protein structural determination, see, e.g., Blundell and Johnson (1976) Protein Crystallography, Academic Press, New York, which is hereby incorporated herein by reference.

II. Activities

20 The cytokine receptor-like proteins will have a number of different biological activities, e.g., modulating cell proliferation, or in phosphate metabolism, being added to or removed from specific substrates, typically proteins. Such will generally result in modulation of an inflammatory function, other innate immunity response, or a morphological effect. The subunit will probably have a specific low affinity binding to the ligand.

30 The DCRS2 has the characteristic motifs of a receptor signaling through the JAK pathway. See, e.g., Ihle, et al. (1997) Stem Cells 15(suppl. 1):105-111; Silvennoinen, et al. (1997) APMIS 105:497-509; Levy (1997) Cytokine Growth Factor Review 8:81-90; Winston and Hunter (1996) Current Biol. 6:668-671; Barrett (1996) Baillieres Clin. Gastroenterol. 10:1-15; and Briscoe, et al. (1996) Philos. Trans. R. Soc. Lond. B. Biol. Sci. 351:167-171.

35 The biological activities of the cytokine receptor subunits will be related to addition or removal of phosphate moieties to substrates, typically in a specific manner, but

5 occasionally in a non specific manner. Substrates may be
identified, or conditions for enzymatic activity may be assayed
by standard methods, e.g., as described in Hardie, et al. (eds.
10 1995) The Protein Kinase FactBook vols. I and II, Academic
5 Press, San Diego, CA; Hanks, et al. (1991) Meth. Enzymol.
200:38-62; Hunter, et al. (1992) Cell 70:375-388; Lewin (1990)
Cell 61:743-752; Pines, et al. (1991) Cold Spring Harbor Symp.
15 Quant. Biol. 56:449-463; and Parker, et al. (1993) Nature
363:736-738.

10 The receptor subunits may combine to form functional
complexes, e.g., which may be useful for binding ligand or
preparing antibodies. These will have substantial diagnostic
20 uses, including detection or quantitation.

15 III. Nucleic Acids

This invention contemplates use of isolated nucleic acid
25 or fragments, e.g., which encode these or closely related
proteins, or fragments thereof, e.g., to encode a corresponding
polypeptide, preferably one which is biologically active. In
30 addition, this invention covers isolated or recombinant DNAs
which encode combinations of such proteins or polypeptides
having characteristic sequences, e.g., of the DCRS2s.
Typically, the nucleic acid is capable of hybridizing, under
appropriate conditions, with a nucleic acid sequence segment
35 shown in Table 1, but preferably not with a corresponding
segment of other receptors described in Table 3. Said
biologically active protein or polypeptide can be a full length
protein, or fragment, and will typically have a segment of
40 amino acid sequence highly homologous, e.g., exhibiting
30 significant stretches of identity, to one shown in Table 1.
Further, this invention covers the use of isolated or
recombinant nucleic acid, or fragments thereof, which encode
45 proteins having fragments which are equivalent to the DCRS2
proteins. The isolated nucleic acids can have the respective
35 regulatory sequences in the 5' and 3' flanks, e.g., promoters,
enhancers, poly-A addition signals, and others from the natural
gene. Combinations, as described, are also provided.
50

5 An "isolated" nucleic acid is a nucleic acid, e.g., an
RNA, DNA, or a mixed polymer, which is substantially pure,
e.g., separated from other components which naturally accompany
10 a native sequence, such as ribosomes, polymerases, and flanking
5 genomic sequences from the originating species. The term
embraces a nucleic acid sequence which has been removed from
its naturally occurring environment, and includes recombinant
or cloned DNA isolates, which are thereby distinguishable from
15 naturally occurring compositions, and chemically synthesized
10 analogs or analogs biologically synthesized by heterologous
systems. A substantially pure molecule includes isolated forms
of the molecule, either completely or substantially pure.

20 An isolated nucleic acid will generally be a homogeneous
composition of molecules, but will, in some embodiments,
15 contain heterogeneity, preferably minor. This heterogeneity is
typically found at the polymer ends or portions not critical to
25 a desired biological function or activity.

A "recombinant" nucleic acid is typically defined either
by its method of production or its structure. In reference to
20 its method of production, e.g., a product made by a process,
30 the process is use of recombinant nucleic acid techniques,
e.g., involving human intervention in the nucleotide sequence.
Typically this intervention involves in vitro manipulation,
although under certain circumstances it may involve more
35 classical animal breeding techniques. Alternatively, it can be
a nucleic acid made by generating a sequence comprising fusion
of two fragments which are not naturally contiguous to each
other, but is meant to exclude products of nature, e.g.,
40 naturally occurring mutants as found in their natural state.
30 Thus, for example, products made by transforming cells with an
unnaturally occurring vector is encompassed, as are nucleic
acids comprising sequence derived using any synthetic
45 oligonucleotide process. Such a process is often done to
replace a codon with a redundant codon encoding the same or a
35 conservative amino acid, while typically introducing or
removing a restriction enzyme sequence recognition site.
50 Alternatively, the process is performed to join together
nucleic acid segments of desired functions to generate a single

5 genetic entity comprising a desired combination of functions
not found in the commonly available natural forms, e.g.,
encoding a fusion protein. Restriction enzyme recognition
10 sites are often the target of such artificial manipulations,
5 but other site specific targets, e.g., promoters, DNA
replication sites, regulation sequences, control sequences, or
other useful features may be incorporated by design. A similar
concept is intended for a recombinant, e.g., fusion,
15 polypeptide. This will include a dimeric repeat. Specifically
10 included are synthetic nucleic acids which, by genetic code
redundancy, encode equivalent polypeptides to fragments of
DCRS2 and fusions of sequences from various different related
20 molecules, e.g., other cytokine receptor family members.

A "fragment" in a nucleic acid context is a contiguous
15 segment of at least about 17 nucleotides, generally at least 21
nucleotides, more generally at least 25 nucleotides, ordinarily
25 at least 30 nucleotides, more ordinarily at least 35
nucleotides, often at least 39 nucleotides, more often at least
45 nucleotides, typically at least 50 nucleotides, more
20 typically at least 55 nucleotides, usually at least 60
30 nucleotides, more usually at least 66 nucleotides, preferably
at least 72 nucleotides, more preferably at least 79
nucleotides, and in particularly preferred embodiments will be
35 at least 85 or more nucleotides. Typically, fragments of
25 different genetic sequences can be compared to one another over
appropriate length stretches, particularly defined segments
such as the domains described below.

A nucleic acid which codes for the DCRS2 will be
40 particularly useful to identify genes, mRNA, and cDNA species
30 which code for itself or closely related proteins, as well as
DNAs which code for polymorphic, allelic, or other genetic
variants, e.g., from different individuals or related species.
45 Preferred probes for such screens are those regions of the
interleukin which are conserved between different polymorphic
35 variants or which contain nucleotides which lack specificity,
and will preferably be full length or nearly so. In other
50 situations, polymorphic variant specific sequences will be more
useful.

5 This invention further covers recombinant nucleic acid
molecules and fragments having a nucleic acid sequence
identical to or highly homologous to the isolated DNA set forth
10 herein. In particular, the sequences will often be operably
5 linked to DNA segments which control transcription,
translation, and DNA replication. These additional segments
typically assist in expression of the desired nucleic acid
segment.

15 Homologous, or highly identical, nucleic acid sequences,
10 when compared to one another, e.g., DCRS2 sequences, exhibit
significant similarity. The standards for homology in nucleic
acids are either measures for homology generally used in the
20 art by sequence comparison or based upon hybridization
conditions. Comparative hybridization conditions are
15 described in greater detail below.

25 Substantial identity in the nucleic acid sequence
comparison context means either that the segments, or their
complementary strands, when compared, are identical when
optimally aligned, with appropriate nucleotide insertions or
20 deletions, in at least about 60% of the nucleotides, generally
at least 66%, ordinarily at least 71%, often at least 76%, more
often at least 80%, usually at least 84%, more usually at least
30 88%, typically at least 91%, more typically at least about 93%,
preferably at least about 95%, more preferably at least about
35 96 to 98% or more, and in particular embodiments, as high at
about 99% or more of the nucleotides, including, e.g., segments
encoding structural domains such as the segments described
below. Alternatively, substantial identity will exist when the
40 segments will hybridize under selective hybridization
30 conditions, to a strand or its complement, typically using a
sequence derived from Table 1. Typically, selective
hybridization will occur when there is at least about 55%
45 homology over a stretch of at least about 14 nucleotides, more
typically at least about 65%, preferably at least about 75%,
35 and more preferably at least about 90%. See, Kanehisa (1984)
Nucl. Acids Res. 12:203-213, which is incorporated herein by
50 reference. The length of homology comparison, as described,
may be over longer stretches, and in certain embodiments will

5 be over a stretch of at least about 17 nucleotides, generally
at least about 20 nucleotides, ordinarily at least about 24
nucleotides, usually at least about 28 nucleotides, typically
10 at least about 32 nucleotides, more typically at least about 40
5 nucleotides, preferably at least about 50 nucleotides, and more
preferably at least about 75 to 100 or more nucleotides. This
includes, e.g., 125, 150, 175, 200, 225, 246, 273, and other
lengths.

15 Stringent conditions, in referring to homology in the
10 hybridization context, will be stringent combined conditions of
salt, temperature, organic solvents, and other parameters
typically controlled in hybridization reactions. Stringent
20 temperature conditions will usually include temperatures in
excess of about 30° C, more usually in excess of about 37° C,
15 typically in excess of about 45° C, more typically in excess of
about 55° C, preferably in excess of about 65° C, and more
25 preferably in excess of about 70° C. Stringent salt conditions
will ordinarily be less than about 500 mM, usually less than
about 400 mM, more usually less than about 300 mM, typically
20 less than about 200 mM, preferably less than about 100 mM, and
30 more preferably less than about 80 mM, even down to less than
about 20 mM. However, the combination of parameters is much
more important than the measure of any single parameter. See,
e.g., Wetmur and Davidson (1968) J. Mol. Biol. 31:349-370,
35 25 which is hereby incorporated herein by reference.

The isolated DNA can be readily modified by nucleotide
substitutions, nucleotide deletions, nucleotide insertions, and
inversions of nucleotide stretches. These modifications result
40 in novel DNA sequences which encode this protein or its
30 derivatives. These modified sequences can be used to produce
mutant proteins (mutans) or to enhance the expression of
variant species. Enhanced expression may involve gene
45 amplification, increased transcription, increased translation,
and other mechanisms. Such mutant DCRS2-like derivatives
35 include predetermined or site-specific mutations of the protein
or its fragments, including silent mutations using genetic code
50 degeneracy. "Mutant DCRS2" as used herein encompasses a
polypeptide otherwise falling within the homology definition of

5 the DCRS2 as set forth above, but having an amino acid sequence
which differs from that of other cytokine receptor-like
10 proteins as found in nature, whether by way of deletion,
substitution, or insertion. In particular, "site specific
5 mutant DCRS2" encompasses a protein having substantial sequence
identity with a protein of Table 1, and typically shares most
of the biological activities or effects of the forms disclosed
herein.

15 Although site specific mutation sites are predetermined,
10 mutants need not be site specific. Mammalian DCRS2 mutagenesis
can be achieved by making amino acid insertions or deletions in
the gene, coupled with expression. Substitutions, deletions,
20 insertions, or many combinations may be generated to arrive at
a final construct. Insertions include amino- or carboxy-
15 terminal fusions. Random mutagenesis can be conducted at a
target codon and the expressed mammalian DCRS2 mutants can then
be screened for the desired activity, providing some aspect of
25 a structure-activity relationship. Methods for making
substitution mutations at predetermined sites in DNA having a
20 known sequence are well known in the art, e.g., by M13 primer
mutagenesis. See also Sambrook, et al. (1989) and Ausubel, et
30 al. (1987 and periodic Supplements).

35 The mutations in the DNA normally should not place coding
sequences out of reading frames and preferably will not create
25 complementary regions that could hybridize to produce secondary
mRNA structure such as loops or hairpins.

40 The phosphoramidite method described by Beaucage and
Carruthers (1981) Tetra. Letts. 22:1859-1862, will produce
suitable synthetic DNA fragments. A double stranded fragment
30 will often be obtained either by synthesizing the complementary
strand and annealing the strand together under appropriate
conditions or by adding the complementary strand using DNA
45 polymerase with an appropriate primer sequence.

35 Polymerase chain reaction (PCR) techniques can often be
applied in mutagenesis. Alternatively, mutagenesis primers are
commonly used methods for generating defined mutations at
50 predetermined sites. See, e.g., Innis, et al. (eds. 1990) PCR
Protocols: A Guide to Methods and Applications Academic Press,

5 San Diego, CA; and Dieffenbach and Dveksler (1995; eds.) PCR
Primer: A Laboratory Manual Cold Spring Harbor Press, CSH, NY.

10 Certain embodiments of the invention are directed to
5 combination compositions comprising the receptor or ligand
sequences described. In other embodiments, functional portions
of the sequences may be joined to encode fusion proteins. In
other forms, variants of the described sequences may be
substituted.

15
10 IV.. Proteins, Peptides

As described above, the present invention encompasses
primate DCRS2, e.g., whose sequences are disclosed in Table 1,
and described above. Allelic and other variants are also
contemplated, including, e.g., fusion proteins combining
15 portions of such sequences with others, including, e.g.,
epitope tags and functional domains.

25 The present invention also provides recombinant proteins,
e.g., heterologous fusion proteins using segments from these
primate or rodent proteins. A heterologous fusion protein is a
20 fusion of proteins or segments which are naturally not normally
fused in the same manner. Thus, the fusion product of a DCRS2
with another cytokine receptor is a continuous protein molecule
having sequences fused in a typical peptide linkage, typically
made as a single translation product and exhibiting properties,
35 e.g., sequence or antigenicity, derived from each source
peptide. A similar concept applies to heterologous nucleic
acid sequences. Combinations of various designated proteins
into complexes are also provided.

40 In addition, new constructs may be made from combining
30 similar functional or structural domains from other related
proteins, e.g., cytokine receptors or Toll-like receptors,
including species variants. For example, ligand-binding or
other segments may be "swapped" between different new fusion
polypeptides or fragments. See, e.g., Cunningham, et al.
45 (1989) Science 243:1330-1336; and O'Dowd, et al. (1988) J.
35 Biol. Chem. 263:15985-15992, each of which is incorporated
herein by reference. Thus, new chimeric polypeptides
50 exhibiting new combinations of specificities will result from

5 the functional linkage of receptor-binding specificities. For
example, the ligand binding domains from other related receptor
molecules may be added or substituted for other domains of this
10 or related proteins. The resulting protein will often have
5 hybrid function and properties. For example, a fusion protein
may include a targeting domain which may serve to provide
sequestering of the fusion protein to a particular subcellular
organelle.

15 Candidate fusion partners and sequences can be selected
10 from various sequence data bases, e.g., GenBank, c/o
IntelliGenetics, Mountain View, CA; and BCG, University of
Wisconsin Biotechnology Computing Group, Madison, WI, which are
20 each incorporated herein by reference. In particular,
combinations of polypeptide sequences provided in Tables 1 and
15 3 are particularly preferred. Variant forms of the proteins
may be substituted in the described combinations.

25 The present invention particularly provides muteins which
bind cytokine-like ligands, and/or which are affected in signal
transduction. Structural alignment of human DCRS2 with other
20 members of the cytokine receptor family show conserved
features/residues. See Table 3. Alignment of the human DCRS2
30 sequence with other members of the cytokine receptor family
indicates various structural and functionally shared features.
See also, Bazan, et al. (1996) Nature 379:591; Lodi, et al.
25 (1994) Science 263:1762-1766; Sayle and Milner-White (1995)
TIBS 20:374-376; and Gronenberg, et al. (1991) Protein
Engineering 4:263-269.

40 Substitutions with either mouse sequences or human
sequences are particularly preferred. Conversely, conservative
30 substitutions away from the ligand binding interaction regions
will probably preserve most signaling activities; and
conservative substitutions away from the intracellular domains
45 will probably preserve most ligand binding properties.

35 "Derivatives" of the primate DCRS2 include amino acid
sequence mutants, glycosylation variants, metabolic derivatives
and covalent or aggregative conjugates with other chemical
50 moieties. Covalent derivatives can be prepared by linkage of
functionalities to groups which are found in the DCRS2 amino

5 acid side chains or at the N- or C- termini, e.g., by means
which are well known in the art. These derivatives can
include, without limitation, aliphatic esters or amides of the
10 carboxyl terminus, or of residues containing carboxyl side
5 chains, O-acyl derivatives of hydroxyl group-containing
residues, and N-acyl derivatives of the amino terminal amino
acid or amino-group containing residues, e.g., lysine or
arginine. Acyl groups are selected from the group of
15 alkyl-moieties, including C3 to C18 normal alkyl, thereby
10 forming alkanoyl aroyl species.

In particular, glycosylation alterations are included,
e.g., made by modifying the glycosylation patterns of a
20 polypeptide during its synthesis and processing, or in further
processing steps. Particularly preferred means for
15 accomplishing this are by exposing the polypeptide to
glycosylating enzymes derived from cells which normally provide
25 such processing, e.g., mammalian glycosylation enzymes.
Deglycosylation enzymes are also contemplated. Also embraced
are versions of the same primary amino acid sequence which have
20 other minor modifications, including phosphorylated amino acid
residues, e.g., phosphotyrosine, phosphoserine, or
30 phosphothreonine.

A major group of derivatives are covalent conjugates of
the receptors or fragments thereof with other proteins of
35 25 polypeptides. These derivatives can be synthesized in
recombinant culture such as N- or C-terminal fusions or by the
use of agents known in the art for their usefulness in
cross-linking proteins through reactive side groups. Preferred
40 derivatization sites with cross-linking agents are at free
30 amino groups, carbohydrate moieties, and cysteine residues.

Fusion polypeptides between the receptors and other
homologous or heterologous proteins are also provided.
45 Homologous polypeptides may be fusions between different
receptors, resulting in, for instance, a hybrid protein
35 exhibiting binding specificity for multiple different cytokine
ligands, or a receptor which may have broadened or weakened
50 specificity of substrate effect. Likewise, heterologous
fusions may be constructed which would exhibit a combination of

5 properties or activities of the derivative proteins. Typical
examples are fusions of a reporter polypeptide, e.g.,
luciferase, with a segment or domain of a receptor, e.g., a
10 ligand-binding segment, so that the presence or location of a
desired ligand may be easily determined. See, e.g., Dull, et
al., U.S. Patent No. 4,859,609, which is hereby incorporated
herein by reference. Other gene fusion partners include
15 glutathione-S-transferase (GST), bacterial β -galactosidase,
trpE, Protein A, β -lactamase, alpha amylase, alcohol
20 dehydrogenase, and yeast alpha mating factor. See, e.g.,
Godowski, et al. (1988) Science 241:812-816. Labeled proteins
will often be substituted in the described combinations of
proteins.

The phosphoramidite method described by Beaucage and
15 Carruthers (1981) Tetra. Letts. 22:1859-1862, will produce
suitable synthetic DNA fragments. A double stranded fragment
will often be obtained either by synthesizing the complementary
25 strand and annealing the strand together under appropriate
conditions or by adding the complementary strand using DNA
20 polymerase with an appropriate primer sequence.

30 Such polypeptides may also have amino acid residues which
have been chemically modified by phosphorylation, sulfonation,
biotinylation, or the addition or removal of other moieties,
particularly those which have molecular shapes similar to
35 phosphate groups. In some embodiments, the modifications will
be useful labeling reagents, or serve as purification targets,
e.g., affinity ligands.

Fusion proteins will typically be made by either
40 recombinant nucleic acid methods or by synthetic polypeptide
30 methods. Techniques for nucleic acid manipulation and
expression are described generally, for example, in Sambrook,
et al. (1989) Molecular Cloning: A Laboratory Manual (2d ed.),
45 Vols. 1-3, Cold Spring Harbor Laboratory, and Ausubel, et al.
(eds. 1987 and periodic supplements) Current Protocols in
35 Molecular Biology, Greene/Wiley, New York, which are each
incorporated herein by reference. Techniques for synthesis of
50 polypeptides are described, for example, in Merrifield (1963)
J. Amer. Chem. Soc. 85:2149-2156; Merrifield (1986) Science

5 232: 341-347; and Atherton, et al. (1989) Solid Phase Peptide
Synthesis: A Practical Approach, IRL Press, Oxford; each of
which is incorporated herein by reference. See also Dawson, et
10 al. (1994) Science 266:776-779 for methods to make larger
5 polypeptides.

This invention also contemplates the use of derivatives of
a DCRS2 other than variations in amino acid sequence or
glycosylation. Such derivatives may involve covalent or
15 aggregative association with chemical moieties. These
10 derivatives generally fall into three classes: (1) salts, (2)
side chain and terminal residue covalent modifications, and (3)
adsorption complexes, for example with cell membranes. Such
20 covalent or aggregative derivatives are useful as immunogens,
as reagents in immunoassays, or in purification methods such as
15 for affinity purification of a receptor or other binding
molecule, e.g., an antibody. For example, a cytokine ligand
25 can be immobilized by covalent bonding to a solid support such
as cyanogen bromide-activated Sepharose, by methods which are
well known in the art, or adsorbed onto polyolefin surfaces,
20 with or without glutaraldehyde cross-linking, for use in the
assay or purification of a cytokine receptor, antibodies, or
30 other similar molecules. The ligand can also be labeled with a
detectable group, for example radioiodinated by the chloramine
T procedure, covalently bound to rare earth chelates, or
25 conjugated to another fluorescent moiety for use in diagnostic
assays.

A combination, e.g., including a DCRS2, of this invention
can be used as an immunogen for the production of antisera or
40 antibodies specific, e.g., capable of distinguishing between
30 other cytokine receptor family members, for the combinations
described. The complexes can be used to screen monoclonal
antibodies or antigen-binding fragments prepared by
45 immunization with various forms of impure preparations
containing the protein. In particular, the term "antibodies"
35 also encompasses antigen binding fragments of natural
antibodies, e.g., Fab, Fab2, Fv, etc. The purified DCRS2 can
50 also be used as a reagent to detect antibodies generated in
response to the presence of elevated levels of expression, or

5 immunological disorders which lead to antibody production to
the endogenous receptor. Additionally, DCRS2 fragments may
also serve as immunogens to produce the antibodies of the
10 present invention, as described immediately below. For
5 example, this invention contemplates antibodies having binding
affinity to or being raised against the amino acid sequences
shown in Table 1, fragments thereof, or various homologous
peptides. In particular, this invention contemplates
15 antibodies having binding affinity to, or having been raised
10 against, specific fragments which are predicted to be, or
actually are, exposed at the exterior protein surface of the
native DCRS2. Complexes of combinations of proteins will also
20 be useful, and antibody preparations thereto can be made.

The blocking of physiological response to the receptor
15 ligands may result from the inhibition of binding of the ligand
to the receptor, likely through competitive inhibition. Thus,
25 in vitro assays of the present invention will often use
antibodies or antigen binding segments of these antibodies, or
fragments attached to solid phase substrates. These assays
20 will also allow for the diagnostic determination of the effects
of either ligand binding region mutations and modifications, or
30 other mutations and modifications, e.g., which affect signaling
or enzymatic function.

This invention also contemplates the use of competitive
35 25 drug screening assays, e.g., where neutralizing antibodies to
the receptor complexes or fragments compete with a test
compound for binding to a ligand or other antibody. In this
manner, the neutralizing antibodies or fragments can be used to
40 detect the presence of a polypeptide which shares one or more
30 binding sites to a receptor and can also be used to occupy
binding sites on a receptor that might otherwise bind a ligand.

45 V. Making Nucleic Acids and Protein

DNA which encodes the protein or fragments thereof can be
35 35 obtained by chemical synthesis, screening cDNA libraries, or by
screening genomic libraries prepared from a wide variety of
cell lines or tissue samples. Natural sequences can be
50 isolated using standard methods and the sequences provided

5 herein, e.g., in Table 1. Other species counterparts can be
identified by hybridization techniques, or by various PCR
10 techniques, combined with or by searching in sequence
databases, e.g., GenBank.

15 This DNA can be expressed in a wide variety of host cells
for the synthesis of a full-length receptor or fragments which
can in turn, for example, be used to generate polyclonal or
monoclonal antibodies; for binding studies; for construction
10 and expression of modified ligand binding or kinase/phosphatase
domains; and for structure/function studies. Variants or
fragments can be expressed in host cells that are transformed
or transfected with appropriate expression vectors. These
20 molecules can be substantially free of protein or cellular
contaminants, other than those derived from the recombinant
15 host, and therefore are particularly useful in pharmaceutical
compositions when combined with a pharmaceutically acceptable
25 carrier and/or diluent. The protein, or portions thereof, may
be expressed as fusions with other proteins. Combinations of
the described proteins, or nucleic acids encoding them, are
20 particularly interesting.

30 Expression vectors are typically self-replicating DNA or
RNA constructs containing the desired receptor gene or its
fragments, usually operably linked to suitable genetic control
elements that are recognized in a suitable host cell. These
35 control elements are capable of effecting expression within a
suitable host. The multiple genes may be coordinately
expressed, and may be on a polycistronic message. The specific
type of control elements necessary to effect expression will
40 depend upon the eventual host cell used. Generally, the
genetic control elements can include a prokaryotic promoter
30 system or a eukaryotic promoter expression control system, and
typically include a transcriptional promoter, an optional
operator to control the onset of transcription, transcription
45 enhancers to elevate the level of mRNA expression, a sequence
that encodes a suitable ribosome binding site, and sequences
35 that terminate transcription and translation. Expression
vectors also usually contain an origin of replication that
50 allows the vector to replicate independently of the host cell.

5 The vectors of this invention include those which contain
DNA which encodes a combination of proteins, as described, or a
biologically active equivalent polypeptide. The DNA can be
10 under the control of a viral promoter and can encode a
5 selection marker. This invention further contemplates use of
such expression vectors which are capable of expressing
eukaryotic cDNAs coding for such proteins in a prokaryotic or
eukaryotic host, where the vector is compatible with the host
15 and where the eukaryotic cDNAs are inserted into the vector
20 such that growth of the host containing the vector expresses
the cDNAs in question. Usually, expression vectors are
designed for stable replication in their host cells or for
20 amplification to greatly increase the total number of copies of
the desirable gene per cell. It is not always necessary to
15 require that an expression vector replicate in a host cell,
e.g., it is possible to effect transient expression of the
25 protein or its fragments in various hosts using vectors that do
not contain a replication origin that is recognized by the host
cell. It is also possible to use vectors that cause
20 integration of the protein encoding portions into the host DNA
by recombination.

30 Vectors, as used herein, comprise plasmids, viruses,
bacteriophage, integratable DNA fragments, and other vehicles
which enable the integration of DNA fragments into the genome
35 of the host. Expression vectors are specialized vectors which
25 contain genetic control elements that effect expression of
operably linked genes. Plasmids are the most commonly used
form of vector but all other forms of vectors which serve an
equivalent function and which are, or become, known in the art
40 are suitable for use herein. See, e.g., Pouwels, et al. (1985
30 and Supplements) Cloning Vectors: A Laboratory Manual,
Elsevier, N.Y., and Rodriguez, et al. (eds. 1988) Vectors: A
45 Survey of Molecular Cloning Vectors and Their Uses,
Buttersworth, Boston, which are incorporated herein by
35 reference.

50 Transformed cells are cells, preferably mammalian, that
have been transformed or transfected with vectors constructed
using recombinant DNA techniques. Transformed host cells

5 usually express the desired proteins, but for purposes of
cloning, amplifying, and manipulating its DNA, do not need to
express the subject proteins. This invention further
10 contemplates culturing transformed cells in a nutrient medium,
5 thus permitting the proteins to accumulate. The proteins can
be recovered, either from the culture or, in certain instances,
from the culture medium.

15 For purposes of this invention, nucleic sequences are
operably linked when they are functionally related to each
10 other. For example, DNA for a presequence or secretory leader
is operably linked to a polypeptide if it is expressed as a
preprotein or participates in directing the polypeptide to the
20 cell membrane or in secretion of the polypeptide. A promoter
is operably linked to a coding sequence if it controls the
15 transcription of the polypeptide; a ribosome binding site is
operably linked to a coding sequence if it is positioned to
25 permit translation. Usually, operably linked means contiguous
and in reading frame, however, certain genetic elements such as
20 repressor genes are not contiguously linked but still bind to
operator sequences that in turn control expression.

30 Suitable host cells include prokaryotes, lower eukaryotes,
and higher eukaryotes. Prokaryotes include both gram negative
and gram positive organisms, e.g., E. coli and B. subtilis.
Lower eukaryotes include yeasts, e.g., S. cerevisiae and
35 Pichia, and species of the genus Dictyostelium. Higher
25 eukaryotes include established tissue culture cell lines from
animal cells, both of non-mammalian origin, e.g., insect cells,
and birds, and of mammalian origin, e.g., human, primates, and
40 rodents.

30 Prokaryotic host-vector systems include a wide variety of
vectors for many different species. As used herein, E. coli
and its vectors will be used generically to include equivalent
45 vectors used in other prokaryotes. A representative vector for
amplifying DNA is pBR322 or many of its derivatives. Vectors
35 that can be used to express the receptor or its fragments
include, but are not limited to, such vectors as those
50 containing the lac promoter (pUC-series); trp promoter
(pBR322-trp); Ipp promoter (the pIN-series); lambda-pP or pR

5 promoters (pOTS); or hybrid promoters such as ptac (pDR540).
See Brosius, et al. (1988) "Expression Vectors Employing
10 Lambda-, trp-, lac-, and Ipp-derived Promoters", in Vectors: A
Survey of Molecular Cloning Vectors and Their Uses, (eds.
5 Rodriguez and Denhardt), Butterworth, Boston, Chapter 10, pp.
205-236, which is incorporated herein by reference.

15 Lower eukaryotes, e.g., yeasts and Dictyostelium, may be
transformed with DCRS2 sequence containing vectors. For
purposes of this invention, the most common lower eukaryotic
10 host is the baker's yeast, Saccharomyces cerevisiae. It will
be used to generically represent lower eukaryotes although a
number of other strains and species are also available. Yeast
20 vectors typically consist of a replication origin (unless of
the integrating type), a selection gene, a promoter, DNA
15 encoding the receptor or its fragments, and sequences for
translation termination, polyadenylation, and transcription
25 termination. Suitable expression vectors for yeast include
such constitutive promoters as 3-phosphoglycerate kinase and
various other glycolytic enzyme gene promoters or such
20 inducible promoters as the alcohol dehydrogenase 2 promoter or
metallothionine promoter. Suitable vectors include derivatives
30 of the following types: self-replicating low copy number (such
as the YRp-series), self-replicating high copy number (such as
the YEp-series); integrating types (such as the YIp-series), or
35 25 mini-chromosomes (such as the YCp-series).

Higher eukaryotic tissue culture cells are normally the
preferred host cells for expression of the functionally active
interleukin or receptor proteins. In principle, many higher
40 eukaryotic tissue culture cell lines are workable, e.g., insect
30 baculovirus expression systems, whether from an invertebrate or
vertebrate source. However, mammalian cells are preferred.
Transformation or transfection and propagation of such cells
45 has become a routine procedure. Examples of useful cell lines
include HeLa cells, Chinese hamster ovary (CHO) cell lines,
35 baby rat kidney (BRK) cell lines, insect cell lines, bird cell
lines, and monkey (COS) cell lines. Expression vectors for
50 such cell lines usually include an origin of replication, a
promoter, a translation initiation site, RNA splice sites (if

5 genomic DNA is used), a polyadenylation site, and a
transcription termination site. These vectors also usually
contain a selection gene or amplification gene. Suitable
10 expression vectors may be plasmids, viruses, or retroviruses
5 carrying promoters derived, e.g., from such sources as from
adenovirus, SV40, parvoviruses, vaccinia virus, or
cytomegalovirus. Representative examples of suitable
expression vectors include pCDNA1; pCD, see Okayama, et al.
15 (1985) Mol. Cell Biol. 5:1136-1142; pMC1neo PolyA, see Thomas,
10 et al. (1987) Cell 51:503-512; and a baculovirus vector such as
pAC 373 or pAC 610.

20 For secreted proteins and some membrane proteins, an open
reading frame usually encodes a polypeptide that consists of a
mature or secreted product covalently linked at its N-terminus
15 to a signal peptide. The signal peptide is cleaved prior to
secretion of the mature, or active, polypeptide. The cleavage
25 site can be predicted with a high degree of accuracy from
empirical rules, e.g., von-Heijne (1986) Nucleic Acids Research
14:4683-4690 and Nielsen, et al. (1997) Protein Eng. 10:1-12,
20 and the precise amino acid composition of the signal peptide
often does not appear to be critical to its function, e.g.,
30 Randall, et al. (1989) Science 243:1156-1159; Kaiser et al.
(1987) Science 235:312-317. The mature proteins of the
invention can be readily determined using standard methods.

35 25 It will often be desired to express these polypeptides in
a system which provides a specific or defined glycosylation
pattern. In this case, the usual pattern will be that provided
naturally by the expression system. However, the pattern will
40 be modifiable by exposing the polypeptide, e.g., an
30 unglycosylated form, to appropriate glycosylating proteins
introduced into a heterologous expression system. For example,
the receptor gene may be co-transformed with one or more genes
45 encoding mammalian or other glycosylating enzymes. Using this
approach, certain mammalian glycosylation patterns will be
35 achievable in prokaryote or other cells. Expression in
prokaryote cells will typically lead to unglycosylated forms of
50 protein.

5 The source of DCRS2 can be a eukaryotic or prokaryotic
host expressing recombinant DCRS2, such as is described above.
The source can also be a cell line, but other mammalian cell
10 lines are also contemplated by this invention, with the
5 preferred cell line being from the human species.

Now that the sequences are known, the primate DCRS2,
fragments, or derivatives thereof can be prepared by
conventional processes for synthesizing peptides. These
15 include processes such as are described in Stewart and Young
10 (1984) Solid Phase Peptide Synthesis, Pierce Chemical Co.,
Rockford, IL; Bodanszky and Bodanszky (1984) The Practice of
Peptide Synthesis, Springer-Verlag, New York; and Bodanszky
20 (1984) The Principles of Peptide Synthesis, Springer-Verlag,
New York; all of each which are incorporated herein by
15 reference. For example, an azide process, an acid chloride
process, an acid anhydride process, a mixed anhydride process,
25 an active ester process (for example, p-nitrophenyl ester,
N-hydroxysuccinimide ester, or cyanomethyl ester), a
carbodiimidazole process, an oxidative-reductive process, or a
20 dicyclohexylcarbodiimide (DCCD)/additive process can be used.
Solid phase and solution phase syntheses are both applicable to
30 the foregoing processes. Similar techniques can be used with
partial DCRS2 sequences.

The DCRS2 proteins, fragments, or derivatives are suitably
35 25 prepared in accordance with the above processes as typically
employed in peptide synthesis, generally either by a so-called
stepwise process which comprises condensing an amino acid to
the terminal amino acid, one by one in sequence, or by coupling
40 peptide fragments to the terminal amino acid. Amino groups
30 that are not being used in the coupling reaction typically must
be protected to prevent coupling at an incorrect location.

If a solid phase synthesis is adopted, the C-terminal
45 amino acid is bound to an insoluble carrier or support through
its carboxyl group. The insoluble carrier is not particularly
35 limited as long as it has a binding capability to a reactive
carboxyl group. Examples of such insoluble carriers include
50 halomethyl resins, such as chloromethyl resin or bromomethyl

5 resin, hydroxymethyl resins, phenol resins,
tert-alkyloxycarbonylhydrazidated resins, and the like.

10 An amino group-protected amino acid is bound in sequence
5 through condensation of its activated carboxyl group and the
reactive amino group of the previously formed peptide or chain,
to synthesize the peptide step by step. After synthesizing the
complete sequence, the peptide is split off from the insoluble
15 carrier to produce the peptide. This solid-phase approach is
generally described by Merrifield, et al. (1963) in J. Am.
20 Chem. Soc. 85:2149-2156, which is incorporated herein by
reference.

20 The prepared protein and fragments thereof can be isolated
and purified from the reaction mixture by means of peptide
separation, for example, by extraction, precipitation,
15 electrophoresis, various forms of chromatography, and the like.
The receptors of this invention can be obtained in varying
25 degrees of purity depending upon desired uses. Purification
can be accomplished by use of the protein purification
techniques disclosed herein, see below, or by the use of the
20 antibodies herein described in methods of immunoabsorbant
affinity chromatography. This immunoabsorbant affinity
30 chromatography is carried out by first linking the antibodies
to a solid support and then contacting the linked antibodies
with solubilized lysates of appropriate cells, lysates of other
35 25 cells expressing the receptor, or lysates or supernatants of
cells producing the protein as a result of DNA techniques, see
below.

40 Generally, the purified protein will be at least about 40%
pure, ordinarily at least about 50% pure, usually at least
30 about 60% pure, typically at least about 70% pure, more
typically at least about 80% pure, preferable at least about
45 90% pure and more preferably at least about 95% pure, and in
particular embodiments, 97%-99% or more. Purity will usually
be on a weight basis, but can also be on a molar basis.
35 Different assays will be applied as appropriate. Individual
proteins may be purified and thereafter combined.

50 VI. Antibodies

5

Antibodies can be raised to the various mammalian, e.g., primate DCRS2 proteins and fragments thereof, both in naturally occurring native forms and in their recombinant forms, the difference being that antibodies to the active receptor are

10

5 more likely to recognize epitopes which are only present in the native conformations. Denatured antigen detection can also be useful in, e.g., Western analysis. Anti-idiotypic antibodies are also contemplated, which would be useful as agonists or

15

antagonists of a natural receptor or an antibody.

10

Antibodies, including binding fragments and single chain versions, against predetermined fragments of the protein can be raised by immunization of animals with conjugates of the fragments with immunogenic proteins. Monoclonal antibodies are prepared from cells secreting the desired antibody. These

20

antibodies can be screened for binding to normal or defective protein, or screened for agonistic or antagonistic activity.

15

These monoclonal antibodies will usually bind with at least a K_D of about 1 mM, more usually at least about 300 μ M, typically at least about 100 μ M, more typically at least about 30 μ M, preferably at least about 10 μ M, and more preferably at least about 3 μ M or better.

25

20

30

The antibodies, including antigen binding fragments, of this invention can have significant diagnostic or therapeutic value. They can be potent antagonists that bind to the

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25

receptor and inhibit binding to ligand or inhibit the ability of the receptor to elicit a biological response, e.g., act on its substrate. They also can be useful as non-neutralizing antibodies and can be coupled to toxins or radionuclides to bind producing cells, or cells localized to the source of the

40

30

interleukin. Further, these antibodies can be conjugated to drugs or other therapeutic agents, either directly or indirectly by means of a linker.

45

The antibodies of this invention can also be useful in diagnostic applications. As capture or non-neutralizing

35

antibodies, they might bind to the receptor without inhibiting ligand or substrate binding. As neutralizing antibodies, they can be useful in competitive binding assays. They will also be useful in detecting or quantifying ligand. They may be used as

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55

5 reagents for Western blot analysis, or for immunoprecipitation
or immunopurification of the respective protein. Likewise,
nucleic acids and proteins may be immobilized to solid
10 substrates for affinity purification or detection methods. The
5 substrates may be, e.g., solid resin beads or sheets of
plastic.

Protein fragments may be joined to other materials,
particularly polypeptides, as fused or covalently joined
15 polypeptides to be used as immunogens. Mammalian cytokine
10 receptors and fragments may be fused or covalently linked to a
variety of immunogens, such as keyhole limpet hemocyanin,
bovine serum albumin, tetanus toxoid, etc. See Microbiology,
20 Hoeber Medical Division, Harper and Row, 1969; Landsteiner
(1962) Specificity of Serological Reactions, Dover
15 Publications, New York; and Williams, et al. (1967) Methods in
Immunology and Immunochemistry, Vol. 1, Academic Press, New
25 York; each of which are incorporated herein by reference, for
descriptions of methods of preparing polyclonal antisera. A
typical method involves hyperimmunization of an animal with an
20 antigen. The blood of the animal is then collected shortly
after the repeated immunizations and the gamma globulin is
30 isolated.

In some instances, it is desirable to prepare monoclonal
antibodies from various mammalian hosts, such as mice, rodents,
35 25 primates, humans, etc. Description of techniques for preparing
such monoclonal antibodies may be found in, e.g., Stites, et
al. (eds.) Basic and Clinical Immunology (4th ed.), Lange
Medical Publications, Los Altos, CA, and references cited
40 therein; Harlow and Lane (1988) Antibodies: A Laboratory
30 Manual, CSH Press; Goding (1986) Monoclonal Antibodies:
Principles and Practice (2d ed.) Academic Press, New York; and
particularly in Kohler and Milstein (1975) in Nature 256:
45 495-497, which discusses one method of generating monoclonal
antibodies. Each of these references is incorporated herein by
35 reference. Summarized briefly, this method involves injecting
an animal with an immunogen. The animal is then sacrificed and
50 cells taken from its spleen, which are then fused with myeloma
cells. The result is a hybrid cell or "hybridoma" that is

5 capable of reproducing in vitro. The population of hybridomas
is then screened to isolate individual clones, each of which
secrete a single antibody species to the immunogen. In this
10 manner, the individual antibody species obtained are the
5 products of immortalized and cloned single B cells from the
immune animal generated in response to a specific site
recognized on the immunogenic substance.

15 Other suitable techniques involve in vitro exposure of
lymphocytes to the antigenic polypeptides or alternatively to
10 selection of libraries of antibodies in phage or similar
vectors. See, Huse, et al. (1989) "Generation of a Large
Combinatorial Library of the Immunoglobulin Repertoire in Phage
20 Lambda," Science 246:1275-1281; and Ward, et al. (1989) Nature
341:544-546, each of which is hereby incorporated herein by
15 reference. The polypeptides and antibodies of the present
invention may be used with or without modification, including
25 chimeric or humanized antibodies. Frequently, the polypeptides
and antibodies will be labeled by joining, either covalently or
non-covalently, a substance which provides for a detectable
20 signal. A wide variety of labels and conjugation techniques
are known and are reported extensively in both the scientific
30 and patent literature. Suitable labels include radionuclides,
enzymes, substrates, cofactors, inhibitors, fluorescent
moieties, chemiluminescent moieties, magnetic particles, and
35 the like. Patents, teaching the use of such labels include
U.S. Patent Nos. 3,817,837; 3,850,752; 3,939,350; 3,996,345;
4,277,437; 4,275,149; and 4,366,241. Also, recombinant or
chimeric immunoglobulins may be produced, see Cabilly, U.S.
40 Patent No. 4,816,567; or made in transgenic mice, see Mendez,
30 et al. (1997) Nature Genetics 15:146-156. These references are
incorporated herein by reference.

45 The antibodies of this invention can also be used for
affinity chromatography in isolating the DCRS2 proteins or
peptides. Columns can be prepared where the antibodies are
35 linked to a solid support, e.g., particles, such as agarose,
Sephadex, or the like, where a cell lysate may be passed
50 through the column, the column washed, followed by increasing
concentrations of a mild denaturant, whereby the purified

5 protein will be released. Alternatively, the protein may be
used to purify antibody. Appropriate cross absorptions or
depletions may be applied.

10 The antibodies may also be used to screen expression
5 libraries for particular expression products. Usually the
antibodies used in such a procedure will be labeled with a
moiety allowing easy detection of presence of antigen by
antibody binding.

15 Antibodies raised against a cytokine receptor will also be
10 used to raise anti-idiotypic antibodies. These will be useful
in detecting or diagnosing various immunological conditions
related to expression of the protein or cells which express the
20 protein. They also will be useful as agonists or antagonists
of the ligand, which may be competitive inhibitors or
15 substitutes for naturally occurring ligands.

25 A cytokine receptor protein that specifically binds to or
that is specifically immunoreactive with an antibody generated
against a defined immunogen, such as an immunogen consisting of
the amino acid sequence of SEQ ID NO: 2, is typically
20 determined in an immunoassay. The immunoassay typically uses a
polyclonal antiserum which was raised, e.g., to a protein of
30 SEQ ID NO: 2. This antiserum is selected to have low
crossreactivity against other cytokine receptor family members,
e.g., IL-11 receptor subunit alpha, IL-6 receptor subunit
35 alpha, or p40, preferably from the same species, and any such
crossreactivity is removed by immunoabsorption prior to use in
the immunoassay.

40 In order to produce antisera for use in an immunoassay,
the protein, e.g., of SEQ ID NO: 2, is isolated as described
30 herein. For example, recombinant protein may be produced in a
mammalian cell line. An appropriate host, e.g., an inbred
strain of mice such as Balb/c, is immunized with the selected
45 protein, typically using a standard adjuvant, such as Freund's
adjuvant, and a standard mouse immunization protocol (see
35 Harlow and Lane, supra). Alternatively, a synthetic peptide
derived from the sequences disclosed herein and conjugated to a
carrier protein can be used an immunogen. Polyclonal sera are
50 collected and titered against the immunogen protein in an

5 immunoassay, e.g., a solid phase immunoassay with the immunogen
immobilized on a solid support. Polyclonal antisera with a
10 titer of 10^4 or greater are selected and tested for their cross
reactivity against other cytokine receptor family members,
5 e.g., IL-11 receptor subunit alpha and/or p40, using a
competitive binding immunoassay such as the one described in
Harlow and Lane, supra, at pages 570-573. Preferably at least
15 two cytokine receptor family members are used in this
determination. These cytokine receptor family members can be
10 produced as recombinant proteins and isolated using standard
molecular biology and protein chemistry techniques as described
herein.

20 Immunoassays in the competitive binding format can be used
for the crossreactivity determinations. For example, the
15 protein of SEQ ID NO: 2 can be immobilized to a solid support.
Proteins added to the assay compete with the binding of the
25 antisera to the immobilized antigen. The ability of the above
proteins to compete with the binding of the antisera to the
immobilized protein is compared to the proteins, e.g., of IL-
20 11 receptor subunit alpha or p40. The percent crossreactivity
for the above proteins is calculated, using standard
30 calculations. Those antisera with less than 10%
crossreactivity with each of the proteins listed above are
selected and pooled. The cross-reacting antibodies are then
35 removed from the pooled antisera by immunoabsorption with the
above-listed proteins.

The immunoabsorbed and pooled antisera are then used in a
competitive binding immunoassay as described above to compare a
40 second protein to the immunogen protein (e.g., the DCRS2 like
30 protein of SEQ ID NO: 2). In order to make this comparison,
the two proteins are each assayed at a wide range of
45 concentrations and the amount of each protein required to
inhibit 50% of the binding of the antisera to the immobilized
protein is determined. If the amount of the second protein
35 required is less than twice the amount of the protein of the
selected protein or proteins that is required, then the second
50 protein is said to specifically bind to an antibody generated
to the immunogen.

5 It is understood that these cytokine receptor proteins are
members of a family of homologous proteins that comprise at
least 6 so far identified genes. For a particular gene
10 product, such as the DCRS2, the term refers not only to the
5 amino acid sequences disclosed herein, but also to other
proteins that are allelic, non-allelic, or species variants.
It is also understood that the terms include nonnatural
15 mutations introduced by deliberate mutation using conventional
recombinant technology such as single site mutation, or by
20 excising short sections of DNA encoding the respective
proteins, or by substituting new amino acids, or adding new
amino acids. Such minor alterations typically will
substantially maintain the immunoidentity of the original
molecule and/or its biological activity. Thus, these
25 alterations include proteins that are specifically
immunoreactive with a designated naturally occurring DCRS2
protein. The biological properties of the altered proteins can
be determined by expressing the protein in an appropriate cell
line and measuring the appropriate effect, e.g., upon
30 transfected lymphocytes. Particular protein modifications
considered minor would include conservative substitution of
amino acids with similar chemical properties, as described
above for the cytokine receptor family as a whole. By aligning
35 a protein optimally with the protein of the cytokine receptors
25 and by using the conventional immunoassays described herein to
determine immunoidentity, one can determine the protein
compositions of the invention.

40 VII. Kits and quantitation

30 Both naturally occurring and recombinant forms of the
cytokine receptor like molecules of this invention are
particularly useful in kits and assay methods. For example,
45 these methods would also be applied to screening for binding
activity, e.g., ligands for these proteins. Several methods of
35 automating assays have been developed in recent years so as to
permit screening of tens of thousands of compounds per year.
See, e.g., a BIOMEK automated workstation, Beckman Instruments,
50 Palo Alto, California, and Fodor, et al. (1991) Science

5 251:767-773, which is incorporated herein by reference. The
latter describes means for testing binding by a plurality of
defined polymers synthesized on a solid substrate. The
10 development of suitable assays to screen for a ligand or
5 agonist/antagonist homologous proteins can be greatly
facilitated by the availability of large amounts of purified,
soluble cytokine receptors in an active state such as is
provided by this invention.

15 Purified DCRS2 can be coated directly onto plates for use
10 in the aforementioned ligand screening techniques. However,
non-neutralizing antibodies to these proteins can be used as
capture antibodies to immobilize the respective receptor on the
20 solid phase, useful, e.g., in diagnostic uses.

25 This invention also contemplates use of DCRS2, fragments
15 thereof, peptides, and their fusion products in a variety of
diagnostic kits and methods for detecting the presence of the
protein or its ligand. Alternatively, or additionally,
antibodies against the molecules may be incorporated into the
kits and methods. Typically the kit will have a compartment
20 containing either a DCRS2 peptide or gene segment or a reagent
which recognizes one or the other. Typically, recognition
30 reagents, in the case of peptide, would be a receptor or
antibody, or in the case of a gene segment, would usually be a
hybridization probe.

35 25 A preferred kit for determining the concentration of DCRS2
in a sample would typically comprise a labeled compound, e.g.,
ligand or antibody, having known binding affinity for DCRS2, a
source of DCRS2 (naturally occurring or recombinant) as a
40 positive control, and a means for separating the bound from
30 free labeled compound, for example a solid phase for
immobilizing the DCRS2 in the test sample. Compartments
containing reagents, and instructions, will normally be
45 provided. Appropriate nucleic acid or protein containing kits
are also provided.

50 35 Antibodies, including antigen binding fragments, specific
for mammalian DCRS2 or a peptide fragment, or receptor
fragments are useful in diagnostic applications to detect the
presence of elevated levels of ligand and/or its fragments.

5 Diagnostic assays may be homogeneous (without a separation step
between free reagent and antibody-antigen complex) or
heterogeneous (with a separation step). Various commercial
10 assays exist, such as radioimmunoassay (RIA), enzyme-linked
5 immunosorbent assay (ELISA), enzyme immunoassay (EIA),
enzyme-multiplied immunoassay technique (EMIT),
substrate-labeled fluorescent immunoassay (SLFIA) and the like.
For example, unlabeled antibodies can be employed by using a
15 second antibody which is labeled and which recognizes the
10 antibody to a cytokine receptor or to a particular fragment
thereof. These assays have also been extensively discussed in
the literature. See, e.g., Harlow and Lane (1988) Antibodies:
20 A Laboratory Manual, CSH., and Coligan (ed. 1991 and periodic
supplements) Current Protocols In Immunology Greene/Wiley, New
15 York.

25 Anti-idiotypic antibodies may have similar use to serve as
agonists or antagonists of cytokine receptors. These should be
useful as therapeutic reagents under appropriate circumstances.

20 Frequently, the reagents for diagnostic assays are
supplied in kits, so as to optimize the sensitivity of the
30 assay. For the subject invention, depending upon the nature of
the assay, the protocol, and the label, either labeled or
unlabeled antibody, or labeled ligand is provided. This is
usually in conjunction with other additives, such as buffers,
35 stabilizers, materials necessary for signal production such as
substrates for enzymes, and the like. Preferably, the kit will
also contain instructions for proper use and disposal of the
contents after use. Typically the kit has compartments for
40 each useful reagent, and will contain instructions for proper
30 use and disposal of reagents. Desirably, the reagents are
provided as a dry lyophilized powder, where the reagents may be
reconstituted in an aqueous medium having appropriate
45 concentrations for performing the assay.

35 The aforementioned constituents of the diagnostic assays
may be used without modification or may be modified in a
variety of ways. For example, labeling may be achieved by
50 covalently or non-covalently joining a moiety which directly or
indirectly provides a detectable signal. In many of these

5 assays, a test compound, cytokine receptor, or antibodies
thereto can be labeled either directly or indirectly.
Possibilities for direct labeling include label groups:
10 radiolabels such as ^{125}I , enzymes (U.S. Pat. No. 3,645,090)
5 such as peroxidase and alkaline phosphatase, and fluorescent
labels (U.S. Pat. No. 3,940,475) capable of monitoring the
change in fluorescence intensity, wavelength shift, or
15 fluorescence polarization. Both of the patents are
incorporated herein by reference. Possibilities for indirect
20 labeling include biotinylation of one constituent followed by
binding to avidin coupled to one of the above label groups.

There are also numerous methods of separating the bound
20 from the free ligand, or alternatively the bound from the free
test compound. The cytokine receptor can be immobilized on
15 various matrixes followed by washing. Suitable matrices
include plastic such as an ELISA plate, filters, and beads.
25 Methods of immobilizing the receptor to a matrix include,
without limitation, direct adhesion to plastic, use of a
capture antibody, chemical coupling, and biotin-avidin. The
20 last step in this approach involves the precipitation of
antibody/antigen complex by any of several methods including
30 those utilizing, e.g., an organic solvent such as polyethylene
glycol or a salt such as ammonium sulfate. Other suitable
separation techniques include, without limitation, the
35 fluorescein antibody magnetizable particle method described in
Rattle, et al. (1984) Clin. Chem. 30(9):1457-1461, and the
double antibody magnetic particle separation as described in
U.S. Pat. No. 4,659,678, each of which is incorporated herein
40 by reference.

30 The methods for linking protein or fragments to various
labels have been extensively reported in the literature and do
not require detailed discussion here. Many of the techniques
45 involve the use of activated carboxyl groups either through the
use of carbodiimide or active esters to form peptide bonds, the
35 formation of thioethers by reaction of a mercapto group with an
activated halogen such as chloroacetyl, or an activated olefin
such as maleimide, for linkage, or the like. Fusion proteins
50 will also find use in these applications.

5 Another diagnostic aspect of this invention involves use
of oligonucleotide or polynucleotide sequences taken from the
sequence of an cytokine receptor. These sequences can be used
10 as probes for detecting levels of the respective cytokine
5 receptor in patients suspected of having an immunological
disorder. The preparation of both RNA and DNA nucleotide
sequences, the labeling of the sequences, and the preferred
size of the sequences has received ample description and
15 discussion in the literature. Normally an oligonucleotide
10 probe should have at least about 14 nucleotides, usually at
least about 18 nucleotides, and the polynucleotide probes may
be up to several kilobases. Various labels may be employed,
20 most commonly radionuclides, particularly ³²P. However, other
techniques may also be employed, such as using biotin modified
15 nucleotides for introduction into a polynucleotide. The biotin
then serves as the site for binding to avidin or antibodies,
25 which may be labeled with a wide variety of labels, such as
radionuclides, fluorescers, enzymes, or the like.
Alternatively, antibodies may be employed which can recognize
20 specific duplexes, including DNA duplexes, RNA duplexes,
DNA-RNA hybrid duplexes, or DNA-protein duplexes. The
30 antibodies in turn may be labeled and the assay carried out
where the duplex is bound to a surface, so that upon the
formation of duplex on the surface, the presence of antibody
35 bound to the duplex can be detected. The use of probes to the
novel anti-sense RNA may be carried out in conventional
techniques such as nucleic acid hybridization, plus and minus
40 screening, recombinational probing, hybrid released translation
(HRT), and hybrid arrested translation (HART). This also
30 includes amplification techniques such as polymerase chain
reaction (PCR).

45 Diagnostic kits which also test for the qualitative or
quantitative presence of other markers are also contemplated.
Diagnosis or prognosis may depend on the combination of
35 multiple indications used as markers. Thus, kits may test for
combinations of markers. See, e.g., Viallet, et al. (1989)
50 Progress in Growth Factor Res. 1:89-97.

VIII. Therapeutic Utility

This invention provides reagents with significant therapeutic value. See, e.g., Levitzki (1996) Curr. Opin. Cell Biol. 8:239-244. The cytokine receptors (naturally occurring or recombinant), fragments thereof, mutein receptors, and antibodies, along with compounds identified as having binding affinity to the receptors or antibodies, should be useful in the treatment of conditions exhibiting abnormal expression of the receptors of their ligands. Such abnormality will typically be manifested by immunological disorders. Additionally, this invention should provide therapeutic value in various diseases or disorders associated with abnormal expression or abnormal triggering of response to the ligand. For example, the IL-1 ligands have been suggested to be involved in morphologic development, e.g., dorso-ventral polarity determination, and immune responses, particularly the primitive innate responses. See, e.g., Sun, et al. (1991) Eur. J. Biochem. 196:247-254; and Hultmark (1994) Nature 367:116-117.

Recombinant cytokine receptors, muteins, agonist or antagonist antibodies thereto, or antibodies can be purified and then administered to a patient. These reagents can be combined for therapeutic use with additional active ingredients, e.g., in conventional pharmaceutically acceptable carriers or diluents, along with physiologically innocuous stabilizers and excipients. These combinations can be sterile, e.g., filtered, and placed into dosage forms as by lyophilization in dosage vials or storage in stabilized aqueous preparations. This invention also contemplates use of antibodies or binding fragments thereof which are not complement binding.

Ligand screening using cytokine receptor or fragments thereof can be performed to identify molecules having binding affinity to the receptors. Subsequent biological assays can then be utilized to determine if a putative ligand can provide competitive binding, which can block intrinsic stimulating activity. Receptor fragments can be used as a blocker or antagonist in that it blocks the activity of ligand. Likewise,

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5 a compound having intrinsic stimulating activity can activate
the receptor and is thus an agonist in that it simulates the
activity of ligand, e.g., inducing signaling. This invention
10 further contemplates the therapeutic use of antibodies to
5 cytokine receptors as antagonists.

The quantities of reagents necessary for effective therapy
will depend upon many different factors, including means of
administration, target site, reagent physiological life,
15 pharmacological life, physiological state of the patient, and
10 other medicants administered. Thus, treatment dosages should
be titrated to optimize safety and efficacy. Typically,
dosages used in vitro may provide useful guidance in the
20 amounts useful for in situ administration of these reagents.
Animal testing of effective doses for treatment of particular
15 disorders will provide further predictive indication of human
dosage. Various considerations are described, e.g., in Gilman,
25 et al. (eds. 1990) Goodman and Gilman's: The Pharmacological
Bases of Therapeutics, 8th Ed., Pergamon Press; and Remington's
Pharmaceutical Sciences, 17th ed. (1990), Mack Publishing Co.,
20 Easton, Penn.; each of which is hereby incorporated herein by
reference. Methods for administration are discussed therein
and below, e.g., for oral, intravenous, intraperitoneal, or
30 intramuscular administration, transdermal diffusion, and
others. Pharmaceutically acceptable carriers will include
35 water, saline, buffers, and other compounds described, e.g., in
the Merck Index, Merck & Co., Rahway, New Jersey. Because of
the likely high affinity binding, or turnover numbers, between
40 a putative ligand and its receptors, low dosages of these
reagents would be initially expected to be effective. And the
30 signaling pathway suggests extremely low amounts of ligand may
have effect. Thus, dosage ranges would ordinarily be expected
to be in amounts lower than 1 mM concentrations, typically less
45 than about 10 μ M concentrations, usually less than about 100
nM, preferably less than about 10 pM (picomolar), and most
35 preferably less than about 1 fM (femtomolar), with an
appropriate carrier. Slow release formulations, or slow
release apparatus will often be utilized for continuous
50 administration.

5 Cytokine receptors, fragments thereof, and antibodies or
its fragments, antagonists, and agonists, may be administered
directly to the host to be treated or, depending on the size of
10 the compounds, it may be desirable to conjugate them to carrier
5 proteins such as ovalbumin or serum albumin prior to their
administration. Therapeutic formulations may be administered
in many conventional dosage formulations. While it is possible
for the active ingredient to be administered alone, it is
15 preferable to present it as a pharmaceutical formulation.
10 Formulations comprise at least one active ingredient, as
defined above, together with one or more acceptable carriers
thereof. Each carrier must be both pharmaceutically and
20 physiologically acceptable in the sense of being compatible
with the other ingredients and not injurious to the patient.
15 Formulations include those suitable for oral, rectal, nasal, or
parenteral (including subcutaneous, intramuscular, intravenous
and intradermal) administration. The formulations may
25 conveniently be presented in unit dosage form and may be
prepared by methods well known in the art of pharmacy. See,
20 e.g., Gilman, et al. (eds. 1990) Goodman and Gilman's: The
Pharmacological Bases of Therapeutics, 8th Ed., Pergamon Press;
30 and Remington's Pharmaceutical Sciences, 17th ed. (1990), Mack
Publishing Co., Easton, Penn.; Avis, et al. (eds. 1993)
Pharmaceutical Dosage Forms: Parenteral Medications Dekker, NY;
35 Lieberman, et al. (eds. 1990) Pharmaceutical Dosage Forms:
Tablets Dekker, NY; and Lieberman, et al. (eds. 1990)
Pharmaceutical Dosage Forms: Disperse Systems Dekker, NY. The
40 therapy of this invention may be combined with or used in
association with other therapeutic agents, particularly
30 agonists or antagonists of other cytokine receptor family
members.

IX. Screening

45 Drug screening using DCRS2 or fragments thereof can be
35 performed to identify compounds having binding affinity to the
receptor subunit, including isolation of associated components.
Subsequent biological assays can then be utilized to determine
50 if the compound has intrinsic stimulating activity and is
therefore a blocker or antagonist in that it blocks the

5 activity of the ligand. Likewise, a compound having intrinsic
stimulating activity can activate the receptor and is thus an
agonist in that it simulates the activity of a cytokine ligand.
10 This invention further contemplates the therapeutic use of
5 antibodies to the receptor as cytokine agonists or antagonists.

Similarly, complexes comprising multiple proteins may be
used to screen for ligands or reagents capable of recognizing
the complex. Most cytokine receptors comprise at least two
15 subunits, which may be the same, or distinct. Alternatively,
10 the transmembrane receptor may bind to a complex comprising a
cytokine-like ligand associated with another soluble protein
serving, e.g., as a second receptor subunit.

20 One method of drug screening utilizes eukaryotic or
prokaryotic host cells which are stably transformed with
15 recombinant DNA molecules expressing the DCRS2 in combination
with another cytokine receptor subunit. Cells may be isolated
25 which express a receptor in isolation from other functional
receptors. Such cells, either in viable or fixed form, can be
used for standard antibody/antigen or ligand/receptor binding
20 assays. See also, Parce, et al. (1989) Science 246:243-247;
and Owicki, et al. (1990) Proc. Nat'l Acad. Sci. USA 87:4007-
30 4011, which describe sensitive methods to detect cellular
responses. Competitive assays are particularly useful, where
the cells (source of putative ligand) are contacted and
35 incubated with a labeled receptor or antibody having known
binding affinity to the ligand, such as ¹²⁵I-antibody, and a
test sample whose binding affinity to the binding composition
is being measured. The bound and free labeled binding
40 compositions are then separated to assess the degree of ligand
30 binding. The amount of test compound bound is inversely
proportional to the amount of labeled receptor binding to the
known source. Many techniques can be used to separate bound
45 from free ligand to assess the degree of ligand binding. This
separation step could typically involve a procedure such as
35 adhesion to filters followed by washing, adhesion to plastic
followed by washing, or centrifugation of the cell membranes.
50 Viable cells could also be used to screen for the effects of
drugs on cytokine mediated functions, e.g., second messenger

5 levels, i.e., Ca⁺⁺; cell proliferation; inositol phosphate pool
changes; and others. Some detection methods allow for
elimination of a separation step, e.g., a proximity sensitive
10 detection system. Calcium sensitive dyes will be useful for
5 detecting Ca⁺⁺ levels, with a fluorimeter or a fluorescence
cell sorting apparatus.

15 X. Ligands

The descriptions of the DCRS2 herein provides means to
10 identify ligands, as described above. Such ligand should bind
specifically to the respective receptor with reasonably high
affinity. Various constructs are made available which allow
20 either labeling of the receptor to detect its ligand. For
example, directly labeling cytokine receptor, fusing onto it
15 markers for secondary labeling, e.g., FLAG or other epitope
tags, etc., will allow detection of receptor. This can be
25 histological, as an affinity method for biochemical
purification, or labeling or selection in an expression cloning
approach. A two-hybrid selection system may also be applied
20 making appropriate constructs with the available cytokine
receptor sequences. See, e.g., Fields and Song (1989) Nature
30 340:245-246.

The broad scope of this invention is best understood with
reference to the following examples, which are not intended to
35 25 limit the inventions to the specific embodiments.

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EXAMPLES

I. General Methods

Some of the standard methods are described or referenced, e.g., in Maniatis, et al. (1982) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor Press; Sambrook, et al. (1989) Molecular Cloning: A Laboratory Manual, (2d ed.), vols. 1-3, CSH Press, NY; or Ausubel, et al. (1987 and Supplements) Current Protocols in Molecular Biology, Greene/Wiley, New York. Methods for protein purification include such methods as ammonium sulfate precipitation, column chromatography, electrophoresis, centrifugation, crystallization, and others. See, e.g., Ausubel, et al. (1987 and periodic supplements); Coligan, et al. (ed. 1996) and periodic supplements, Current Protocols In Protein Science Greene/Wiley, New York; Deutscher (1990) "Guide to Protein Purification" in Methods in Enzymology, vol. 182, and other volumes in this series; and manufacturer's literature on use of protein purification products, e.g., Pharmacia, Piscataway, N.J., or Bio-Rad, Richmond, CA. Combination with recombinant techniques allow fusion to appropriate segments, e.g., to a FLAG sequence or an equivalent which can be fused via a protease-removable sequence. See, e.g., Hochuli (1990) "Purification of Recombinant Proteins with Metal Chelate Absorbent" in Setlow (ed.) Genetic Engineering, Principle and Methods 12:87-98, Plenum Press, N.Y.; and Crowe, et al. (1992) QIAexpress: The High Level Expression & Protein Purification System QUIAGEN, Inc., Chatsworth, CA.

Computer sequence analysis is performed, e.g., using available software programs, including those from the GCG (U. Wisconsin) and GenBank sources. Public sequence databases were also used, e.g., from GenBank and others.

Many techniques applicable to IL-10 receptors may be applied to the DCRS2, as described, e.g., in USSN 08/110,683 (IL-10 receptor), which is incorporated herein by reference.

II. Computational Analysis

Human sequences related to cytokine receptors were identified from genomic sequence database using, e.g., the

5 BLAST server (Altschul, et al. (1994) Nature Genet. 6:119-129).
Standard analysis programs may be used to evaluate structure,
e.g., PHD (Rost and Sander (1994) Proteins 19:55-72) and DSC
10 (King and Sternberg (1996) Protein Sci. 5:2298-2310). Standard
5 comparison software includes, e.g., Altschul, et al. (1990) J.
Mol. Biol. 215:403-10; Waterman (1995) Introduction to
Computational Biology: Maps, Sequences, and Genomes Chapman &
Hall; Lander and Waterman (eds. 1995) Calculating the Secrets
15 of Life: Applications of the Mathematical Sciences in Molecular
Biology National Academy Press; and Speed and Waterman (eds.
10 1996) Genetic Mapping and DNA Sequencing (IMA Volumes in
Mathematics and Its Applications, Vol 81) Springer Verlag.

20 III. Cloning of full-length DCRS2 cDNAs; Chromosomal
15 localization

PCR primers derived from the DCRS2 sequence are used to
probe a human cDNA library. Sequences may be derived, e.g.,
25 from Table 1, preferably those adjacent the ends of sequences.
Full length cDNAs for primate, rodent, or other species DCRS2
20 are cloned, e.g., by DNA hybridization screening of λ gt10
phage. PCR reactions are conducted using *T. aquaticus* Taqplus
DNA polymerase (Stratagene) under appropriate conditions.

Chromosome spreads are prepared. In situ hybridization is
performed on chromosome preparations obtained from
25 phytohemagglutinin-stimulated human lymphocytes cultured for 72
h. 5-bromodeoxyuridine was added for the final seven hours of
35 culture (60 μ g/ml of medium), to ensure a posthybridization
chromosomal banding of good quality.

A PCR fragment, amplified with the help of primers, is
40 30 cloned into an appropriate vector. The vector is labeled by
nick-translation with 3 H. The radiolabeled probe is hybridized
to metaphase spreads at final concentration of 200 ng/ml of
hybridization solution as described in Mattei, et al. (1985)
45 Hum. Genet. 69:327-331.

35 After coating with nuclear track emulsion (KODAK NTB₂),
slides are exposed. To avoid any slipping of silver grains
during the banding procedure, chromosome spreads are first
50 stained with buffered Giemsa solution and metaphase
photographed. R-banding is then performed by the fluorochrome-

5 photolysis-Giemsa (FPG) method and metaphases rephotographed before analysis.

Similar appropriate methods are used for other species.

10 5 IV. Localization of DCRS2 mRNA

Human multiple tissue (Cat# 1, 2) and cancer cell line blots (Cat# 7757-1), containing approximately 2 µg of poly(A)⁺ RNA per lane, are purchased from Clontech (Palo Alto, CA).
15 Probes are radiolabeled with [α -³²P] dATP, e.g., using the Amersham Rediprime random primer labeling kit (RPN1633).
20 Prehybridization and hybridizations are performed, e.g., at 65° C in 0.5 M Na₂HPO₄, 7% SDS, 0.5 M EDTA (pH 8.0). High stringency washes are conducted, e.g., at 65° C with two initial washes in 2 x SSC, 0.1% SDS for 40 min followed by a
25 subsequent wash in 0.1 x SSC, 0.1% SDS for 20 min. Membranes are then exposed at -70° C to X-Ray film (Kodak) in the presence of intensifying screens. More detailed studies by cDNA library Southern blots are performed with selected appropriate human DCRS2 clones to examine their expression in hemopoietic
30 or other cell subsets.

Alternatively, two appropriate primers are selected from Table 1. RT-PCR is used on an appropriate mRNA sample selected for the presence of message to produce a cDNA, e.g., a sample which expresses the gene.

35 25 Full length clones may be isolated by hybridization of cDNA libraries from appropriate tissues pre-selected by PCR signal. Northern blots can be performed.

40 30 Message for genes encoding DCRS2 will be assayed by appropriate technology, e.g., PCR, immunoassay, hybridization, or otherwise. Tissue and organ cDNA preparations are available, e.g., from Clontech, Mountain View, CA. Identification of sources of natural expression are useful, as
45 described. And the identification of functional receptor subunit pairings will allow for prediction of what cells
50 35 express the combination of receptor subunits which will result in a physiological responsiveness to each of the cytokine ligands.

5 For mouse distribution, e.g., Southern Analysis can be
performed: DNA (5 µg) from a primary amplified cDNA library was
digested with appropriate restriction enzymes to release the
10 inserts, run on a 1% agarose gel and transferred to a nylon
5 membrane (Schleicher and Schuell, Keene, NH).

Samples for mouse mRNA isolation may include: resting
mouse fibroblastic L cell line (C200); Braf:ER (Braf fusion to
estrogen receptor) transfected cells, control (C201); T cells,
15 TH1 polarized (Mel14 bright, CD4+ cells from spleen, polarized
10 for 7 days with IFN-γ and anti IL-4; T200); T cells, TH2
polarized (Mel14 bright, CD4+ cells from spleen, polarized for
7 days with IL-4 and anti-IFN-γ; T201); T cells, highly TH1
20 polarized (see Openshaw, et al. (1995) J. Exp. Med. 182:1357-
1367; activated with anti-CD3 for 2, 6, 16 h pooled; T202); T
15 cells, highly TH2 polarized (see Openshaw, et al. (1995) J.
Exp. Med. 182:1357-1367; activated with anti-CD3 for 2, 6, 16 h
25 pooled; T203); CD44- CD25+ pre T cells, sorted from thymus
(T204); TH1 T cell clone D1.1, resting for 3 weeks after last
stimulation with antigen (T205); TH1 T cell clone D1.1, 10
20 µg/ml ConA stimulated 15 h (T206); TH2 T cell clone CDC35,
30 resting for 3 weeks after last stimulation with antigen (T207);
TH2 T cell clone CDC35, 10 µg/ml ConA stimulated 15 h (T208);
Mel14+ naive T cells from spleen, resting (T209); Mel14+ T
cells, polarized to Th1 with IFN-γ/IL-12/anti-IL-4 for 6, 12,
35 24 h pooled (T210); Mel14+ T cells, polarized to Th2 with IL-
4/anti-IFN-γ for 6, 13, 24 h pooled (T211); unstimulated mature
B cell leukemia cell line A20 (B200); unstimulated B cell line
CH12 (B201); unstimulated large B cells from spleen (B202); B
40 cells from total spleen, LPS activated (B203); metrizamide
30 enriched dendritic cells from spleen, resting (D200); dendritic
cells from bone marrow, resting (D201); monocyte cell line RAW
264.7 activated with LPS 4 h (M200); bone-marrow macrophages
45 derived with GM and M-CSF (M201); macrophage cell line J774,
resting (M202); macrophage cell line J774 + LPS + anti-IL-10 at
35 0.5, 1, 3, 6, 12 h pooled (M203); macrophage cell line J774 +
LPS + IL-10 at 0.5, 1, 3, 5, 12 h pooled (M204); aerosol
50 challenged mouse lung tissue, Th2 primers, aerosol OVA
challenge 7, 14, 23 h pooled (see Garlisi, et al. (1995)

5 Clinical Immunology and Immunopathology 75:75-83; X206);
Nippostrongylus-infected lung tissue (see Coffman, et al.
10 (1989) Science 245:308-310; X200); total adult lung, normal
(O200); total lung, rag-1 (see Schwarz, et al. (1993)
5 Immunodeficiency 4:249-252; O205); IL-10 K.O. spleen (see Kuhn,
et al. (1991) Cell 75:263-274; X201); total adult spleen,
normal (O201); total spleen, rag-1 (O207); IL-10 K.O. Peyer's
15 patches (O202); total Peyer's patches, normal (O210); IL-10
K.O. mesenteric lymph nodes (X203); total mesenteric lymph
20 nodes, normal (O211); IL-10 K.O. colon (X203); total colon,
normal (O212); NOD mouse pancreas (see Makino, et al. (1980)
Jikken Dobutsu 29:1-13; X205); total thymus, rag-1 (O208);
total kidney, rag-1 (O209); total heart, rag-1 (O202); total
brain, rag-1 (O203); total testes, rag-1 (O204); total liver,
15 rag-1 (O206); rat normal joint tissue (O300); and rat arthritic
joint tissue (X300).

25 Samples for human mRNA isolation may include: peripheral
blood mononuclear cells (monocytes, T cells, NK cells,
granulocytes, B cells), resting (T100); peripheral blood
20 mononuclear cells, activated with anti-CD3 for 2, 6, 12 h
pooled (T101); T cell, TH0 clone Mot 72, resting (T102); T
30 cell, TH0 clone Mot 72, activated with anti-CD28 and anti-CD3
for 3, 6, 12 h pooled (T103); T cell, TH0 clone Mot 72, anergic
treated with specific peptide for 2, 7, 12 h pooled (T104); T
35 cell, TH1 clone HY06, resting (T107); T cell, TH1 clone HY06,
activated with anti-CD28 and anti-CD3 for 3, 6, 12 h pooled
(T108); T cell, TH1 clone HY06, anergic treated with specific
peptide for 2, 6, 12 h pooled (T109); T cell, TH2 clone HY935,
40 resting (T110); T cell, TH2 clone HY935, activated with anti-
30 CD28 and anti-CD3 for 2, 7, 12 h pooled (T111); T cells
CD4+CD45RO- T cells polarized 27 days in anti-CD28, IL-4, and
anti IFN- γ , TH2 polarized, activated with anti-CD3 and anti-
45 CD28 4 h (T116); T cell tumor lines Jurkat and Hut78, resting
(T117); T cell clones, pooled AD130.2, Tc783.12, Tc783.13,
35 Tc783.58, Tc782.69, resting (T118); T cell random $\gamma\delta$ T cell
clones, resting (T119); Splenocytes, resting (B100);
Splenocytes, activated with anti-CD40 and IL-4 (B101); B cell
50 EBV lines pooled WT49, RSB, JY, CVIR, 721.221, RM3, HSY,

5 resting (B102); B cell line JY, activated with PMA and
ionomycin for 1, 6 h pooled (B103); NK 20 clones pooled,
resting (K100); NK 20 clones pooled, activated with PMA and
10 ionomycin for 6 h (K101); NKL clone, derived from peripheral
5 blood of LGL leukemia patient, IL-2 treated (K106); NK
cytotoxic clone 640-A30-1, resting (K107); hematopoietic
precursor line TF1, activated with PMA and ionomycin for 1, 6 h
pooled (C100); U937 premonocytic line, resting (M100); U937
15 premonocytic line, activated with PMA and ionomycin for 1, 6 h
10 pooled (M101); elutriated monocytes, activated with LPS, IFN γ ,
anti-IL-10 for 1, 2, 6, 12, 24 h pooled (M102); elutriated
monocytes, activated with LPS, IFN γ , IL-10 for 1, 2, 6, 12, 24
20 h pooled (M103); elutriated monocytes, activated with LPS,
IFN γ , anti-IL-10 for 4, 16 h pooled (M106); elutriated
15 monocytes, activated with LPS, IFN γ , IL-10 for 4, 16 h pooled
(M107); elutriated monocytes, activated LPS for 1 h (M108);
25 elutriated monocytes, activated LPS for 6 h (M109); DC 70%
CD1a+, from CD34+ GM-CSF, TNF α 12 days, resting (D101); DC 70%
CD1a+, from CD34+ GM-CSF, TNF α 12 days, activated with PMA and
20 ionomycin for 1 hr (D102); DC 70% CD1a+, from CD34+ GM-CSF,
TNF α 12 days, activated with PMA and ionomycin for 6 hr
30 (D103); DC 95% CD1a+, from CD34+ GM-CSF, TNF α 12 days FACS
sorted, activated with PMA and ionomycin for 1, 6 h pooled
(D104); DC 95% CD14+, ex CD34+ GM-CSF, TNF α 12 days FACS
35 sorted, activated with PMA and ionomycin 1, 6 hr pooled (D105);
DC CD1a+ CD86+, from CD34+ GM-CSF, TNF α 12 days FACS sorted,
activated with PMA and ionomycin for 1, 6 h pooled (D106); DC
from monocytes GM-CSF, IL-4 5 days, resting (D107); DC from
40 monocytes GM-CSF, IL-4 5 days, resting (D108); DC from
30 monocytes GM-CSF, IL-4 5 days, activated LPS 4, 16 h pooled
(D109); DC from monocytes GM-CSF, IL-4 5 days, activated TNF α ,
monocyte supe for 4, 16 h pooled (D110); leiomyoma L11 benign
45 tumor (X101); normal myometrium M5 (O115); malignant
leiomyosarcoma GS1 (X103); lung fibroblast sarcoma line MRC5,
35 activated with PMA and ionomycin for 1, 6 h pooled (C101);
kidney epithelial carcinoma cell line CHA, activated with PMA
and ionomycin for 1, 6 h pooled (C102); kidney fetal 28 wk male
50 (O100); lung fetal 28 wk male (O101); liver fetal 28 wk male

5 (O102); heart fetal 28 wk male (O103); brain fetal 28 wk male
(O104); gallbladder fetal 28 wk male (O106); small intestine
fetal 28 wk male (O107); adipose tissue fetal 28 wk male
10 (O108); ovary fetal 25 wk female (O109); uterus fetal 25 wk
5 female (O110); testes fetal 28 wk male (O111); spleen fetal 28
wk male (O112); adult placenta 28 wk (O113); and tonsil
inflamed, from 12 year old (X100).

15 Similar samples may isolated in other species for
evaluation.

10 V. Cloning of species counterparts of DCRS2

20 Various strategies are used to obtain species counterparts
of the DCRS2, preferably from other primates or rodents. One
method is by cross hybridization using closely related species
15 DNA probes. It may be useful to go into evolutionarily similar
species as intermediate steps. Another method is by using
25 specific PCR primers based on the identification of blocks of
similarity or difference between genes, e.g., areas of highly
conserved or nonconserved polypeptide or nucleotide sequence.

20 VI. Production of mammalian DCRS2 protein

30 An appropriate, e.g., GST, fusion construct is engineered
for expression, e.g., in *E. coli*. For example, a mouse IGIF
pGex plasmid is constructed and transformed into *E. coli*.

35 25 Freshly transformed cells are grown, e.g., in LB medium
containing 50 µg/ml ampicillin and induced with IPTG (Sigma,
St. Louis, MO). After overnight induction, the bacteria are
harvested and the pellets containing the DCRS2 protein are
40 isolated. The pellets are homogenized, e.g., in TE buffer (50
30 mM Tris-base pH 8.0, 10 mM EDTA and 2 mM pefabloc) in 2 liters.
This material is passed through a microfluidizer
(Microfluidics, Newton, MA) three times. The fluidized
45 supernatant is spun down on a Sorvall GS-3 rotor for 1 h at
13,000 rpm. The resulting supernatant containing the cytokine
35 receptor protein is filtered and passed over a glutathione-
SEPHAROSE column equilibrated in 50 mM Tris-base pH 8.0. The
50 fractions containing the DCRS2-GST fusion protein are pooled
and cleaved, e.g., with thrombin (Enzyme Research Laboratories,

5 Inc., South Bend, IN). The cleaved pool is then passed over a
Q-SEPHAROSE column equilibrated in 50 mM Tris-base. Fractions
10 containing DCRS2 are pooled and diluted in cold distilled H₂O,
to lower the conductivity, and passed back over a fresh Q-
5 Sepharose column, alone or in succession with an immunoaffinity
antibody column. Fractions containing the DCRS2 protein are
pooled, aliquoted, and stored in the -70° C freezer.

15 Comparison of the CD spectrum with cytokine receptor
protein may suggest that the protein is correctly folded. See
10 Hazuda, et al. (1969) J. Biol. Chem. 264:1689-1693.

VII. Preparation of antibodies specific for DCRS2

20 Inbred Balb/c mice are immunized intraperitoneally with
recombinant forms of the protein, e.g., purified DCRS2 or
15 stable transfected NIH-3T3 cells. Animals are boosted at
appropriate time points with protein, with or without
25 additional adjuvant, to further stimulate antibody production.
Serum is collected, or hybridomas produced with harvested
splensens.

20 Alternatively, Balb/c mice are immunized with cells
30 transformed with the gene or fragments thereof, either
endogenous or exogenous cells, or with isolated membranes
enriched for expression of the antigen. Serum is collected at
the appropriate time, typically after numerous further
35 administrations. Various gene therapy techniques may be
useful, e.g., in producing protein in situ, for generating an
immune response. Serum or antibody preparations may be cross-
absorbed or immunoselected to prepare substantially purified
40 antibodies of defined specificity and high affinity.

30 Monoclonal antibodies may be made. For example,
splenocytes are fused with an appropriate fusion partner and
hybridomas are selected in growth medium by standard
45 procedures. Hybridoma supernatants are screened for the
presence of antibodies which bind to the DCRS2, e.g., by ELISA
35 or other assay. Antibodies which specifically recognize
specific DCRS2 embodiments may also be selected or prepared.

50 In another method, synthetic peptides or purified protein
are presented to an immune system to generate monoclonal or

5 polyclonal antibodies. See, e.g., Coligan (ed. 1991) Current
10 Protocols in Immunology Wiley/Greene; and Harlow and Lane
5 (1989) Antibodies: A Laboratory Manual Cold Spring Harbor
Press. In appropriate situations, the binding reagent is
15 either labeled as described above, e.g., fluorescence or
otherwise, or immobilized to a substrate for panning methods.
Nucleic acids may also be introduced into cells in an animal to
20 produce the antigen, which serves to elicit an immune response.
See, e.g., Wang, et al. (1993) Proc. Nat'l. Acad. Sci. 90:4156-
10 4160; Barry, et al. (1994) BioTechniques 16:616-619; and Xiang,
et al. (1995) Immunity 2: 129-135.

20 VIII. Production of fusion proteins with DCRS2

Various fusion constructs are made with DCRS2. A portion
15 of the appropriate gene is fused to an epitope tag, e.g., a
FLAG tag, or to a two hybrid system construct. See, e.g.,
25 Fields and Song (1989) Nature 340:245-246.

The epitope tag may be used in an expression cloning
procedure with detection with anti-FLAG antibodies to detect a
20 binding partner, e.g., ligand for the respective cytokine
receptor. The two hybrid system may also be used to isolate
30 proteins which specifically bind to DCRS2.

35 IX. Structure activity relationship

25 Information on the criticality of particular residues is
determined using standard procedures and analysis. Standard
mutagenesis analysis is performed, e.g., by generating many
different variants at determined positions, e.g., at the
40 positions identified above, and evaluating biological
30 activities of the variants. This may be performed to the
extent of determining positions which modify activity, or to
focus on specific positions to determine the residues which can
45 be substituted to either retain, block, or modulate biological
activity.

35 Alternatively, analysis of natural variants can indicate
what positions tolerate natural mutations. This may result
50 from populational analysis of variation among individuals, or
across strains or species. Samples from selected individuals

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are analyzed, e.g., by PCR analysis and sequencing. This allows evaluation of population polymorphisms.

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X. Isolation of a ligand for DCRS2

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A cytokine receptor can be used as a specific binding reagent to identify its binding partner, by taking advantage of its specificity of binding, much like an antibody would be used. The binding receptor may be a heterodimer of receptor subunits; or may involve, e.g., a complex of the DCRS2 with another subunit. A binding reagent is either labeled as described above, e.g., fluorescence or otherwise, or immobilized to a substrate for panning methods.

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The binding composition is used to screen an expression library made from a cell line which expresses a binding partner, i.e., ligand, preferably membrane associated. Standard staining techniques are used to detect or sort surface expressed ligand, or surface expressing transformed cells are screened by panning. Screening of intracellular expression is performed by various staining or immunofluorescence procedures. See also McMahan, et al. (1991) EMBO J. 10:2821-2832.

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For example, on day 0, precoat 2-chamber permanox slides with 1 ml per chamber of fibronectin, 10 ng/ml in PBS, for 30 min at room temperature. Rinse once with PBS. Then plate COS cells at 2-3 x 10⁵ cells per chamber in 1.5 ml of growth media. Incubate overnight at 37° C.

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On day 1 for each sample, prepare 0.5 ml of a solution of 66 µg/ml DEAE-dextran, 66 µM chloroquine, and 4 µg DNA in serum free DME. For each set, a positive control is prepared, e.g., of DCRS2-FLAG cDNA at 1 and 1/200 dilution, and a negative mock. Rinse cells with serum free DME. Add the DNA solution and incubate 5 hr at 37° C. Remove the medium and add 0.5 ml 10% DMSO in DME for 2.5 min. Remove and wash once with DME. Add 1.5 ml growth medium and incubate overnight.

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On day 2, change the medium. On days 3 or 4, the cells are fixed and stained. Rinse the cells twice with Hank's Buffered Saline Solution (HBSS) and fix in 4% paraformaldehyde (PFA)/glucose for 5 min. Wash 3X with HBSS. The slides may be stored at -80° C after all liquid is removed. For each chamber, 0.5 ml incubations are performed as follows. Add HBSS/saponin (0.1%) with 32 µl/ml of 1 M NaN₃ for 20 min. Cells are then washed with HBSS/saponin 1X. Add appropriate DCRS2 or

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5 DCRS2/antibody complex to cells and incubate for 30 min. Wash
cells twice with HBSS/saponin. If appropriate, add first
antibody for 30 min. Add second antibody, e.g., Vector anti-
10 mouse antibody, at 1/200 dilution, and incubate for 30 min.
5 Prepare ELISA solution, e.g., Vector Elite ABC horseradish
peroxidase solution, and preincubate for 30 min. Use, e.g., 1
drop of solution A (avidin) and 1 drop solution B (biotin) per
2.5 ml HBSS/saponin. Wash cells twice with HBSS/saponin. Add
15 ABC HRP solution and incubate for 30 min. Wash cells twice
10 with HBSS, second wash for 2 min, which closes cells. Then add
Vector diaminobenzoic acid (DAB) for 5 to 10 min. Use 2 drops
of buffer plus 4 drops DAB plus 2 drops of H₂O₂ per 5 ml of
20 glass distilled water. Carefully remove chamber and rinse
slide in water. Air dry for a few minutes, then add 1 drop of
15 Crystal Mount and a cover slip. Bake for 5 min at 85-90° C.
Evaluate positive staining of pools and progressively
25 subclone to isolation of single genes responsible for the
binding.

Alternatively, receptor reagents are used to affinity
20 purify or sort out cells expressing a putative ligand. See,
e.g., Sambrook, et al. or Ausubel, et al.

Another strategy is to screen for a membrane bound
receptor by panning. The receptor cDNA is constructed as
described above. Immobilization may be achieved by use of
35 appropriate antibodies which recognize, e.g., a FLAG sequence
25 of a DCRS2 fusion construct, or by use of antibodies raised
against the first antibodies. Recursive cycles of selection
and amplification lead to enrichment of appropriate clones and
40 eventual isolation of receptor expressing clones.

30 Phage expression libraries can be screened by mammalian
DCRS2. Appropriate label techniques, e.g., anti-FLAG
antibodies, will allow specific labeling of appropriate clones.

45 All citations herein are incorporated herein by reference to
35 the same extent as if each individual publication or patent
application was specifically and individually indicated to be
50 incorporated by reference.

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Many modifications and variations of this invention can be made without departing from its spirit and scope, as will be apparent to those skilled in the art. The specific embodiments described herein are offered by way of example only, and the invention is to be limited by the terms of the appended claims, along with the full scope of equivalents to which such claims are entitled; and the invention is not to be limited by the specific embodiments that have been presented herein by way of example.

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Claims

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WHAT IS CLAIMED IS:

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1. A composition of matter selected from:
 - a) a substantially pure or recombinant DCRS2 polypeptide comprising at least three distinct nonoverlapping segments of at least four amino acids identical to segments of SEQ ID NO: 2;
 - b) a substantially pure or recombinant DCRS2 polypeptide comprising at least two distinct nonoverlapping segments of at least five amino acids identical to segments of SEQ ID NO: 2;
 - c) a natural sequence DCRS2 comprising mature SEQ ID NO: 2; or
 - d) a fusion polypeptide comprising DCRS2 sequence.

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2. The substantially pure or isolated antigenic DCRS2 polypeptide of Claim 1, wherein said distinct nonoverlapping segments of identity:

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- a) include one of at least eight amino acids;
- b) include one of at least four amino acids and a second of at least five amino acids;
- c) include at least three segments of at least four, five, and six amino acids, or
- d) include one of at least twelve amino acids.

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3. The composition of matter of Claim 1, wherein said:
 - a) DCRS2 polypeptide:
 - i) comprises a mature sequence of Table 1;
 - ii) is an unglycosylated form of DCRS2;
 - iii) is from a primate, such as a human;
 - iv) comprises at least seventeen amino acids of SEQ ID NO: 2;
 - v) exhibits at least four nonoverlapping segments of at least seven amino acids of SEQ ID NO: 2;
 - vi) is a natural allelic variant of DCRS2;
 - vii) has a length at least about 30 amino acids;
 - viii) exhibits at least two non-overlapping epitopes which are specific for a primate DCRS2;
 - ix) is glycosylated;

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x) has a molecular weight of at least 30 kD with natural glycosylation;

xi) is a synthetic polypeptide;

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xii) is attached to a solid substrate;

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xiii) is conjugated to another chemical moiety;

xiv) is a 5-fold or less substitution from natural sequence; or

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xv) is a deletion or insertion variant from a natural sequence.

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4. A composition comprising:

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a) a substantially pure DCRS2 and another cytokine receptor family member;

b) a sterile DCRS2 polypeptide of Claim 1;

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c) said DCRS2 polypeptide of Claim 1 and a carrier, wherein said carrier is:

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i) an aqueous compound, including water, saline, and/or buffer; and/or

ii) formulated for oral, rectal, nasal, topical, or parenteral administration.

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5. The fusion polypeptide of Claim 1, comprising:

a) mature protein sequence of Table 1;

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b) a detection or purification tag, including a FLAG, His6, or Ig sequence; or

c) sequence of another cytokine receptor protein.

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6. A kit comprising a polypeptide of Claim 1, and:

a) a compartment comprising said protein or polypeptide; or

b) instructions for use or disposal of reagents in said kit.

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7. A binding compound comprising an antigen binding site from an antibody, which specifically binds to a natural DCRS2 polypeptide of Claim 1, wherein:

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a) said binding compound is in a container;

b) said DCRS2 polypeptide is from a human;

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- c) said binding compound is an Fv, Fab, or Fab2 fragment;
- d) said binding compound is conjugated to another chemical moiety; or
- e) said antibody:

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- 5 i) is raised against a peptide sequence of a mature polypeptide of Table 1;
- ii) is raised against a mature DCRS2;
- 15 iii) is raised to a purified human DCRS2;
- iv) is immunoselected;
- 10 v) is a polyclonal antibody;
- vi) binds to a denatured DCRS2;
- vii) exhibits a Kd to antigen of at least 30 μ M;
- 20 viii) is attached to a solid substrate, including a bead or plastic membrane;
- 15 ix) is in a sterile composition; or
- x) is detectably labeled, including a radioactive or
- 25 fluorescent label.

8. A kit comprising said binding compound of Claim 7,

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- a) a compartment comprising said binding compound; or
- b) instructions for use or disposal of reagents in said kit.

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25 9. A method of producing an antigen:antibody complex, comprising contacting under appropriate conditions a primate DCRS2 polypeptide with an antibody of Claim 7, thereby allowing said complex to form.

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10. The method of Claim 9, wherein:

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- a) said complex is purified from other cytokine receptors;
- b) said complex is purified from other antibody;
- c) said contacting is with a sample comprising an interferon;
- d) said contacting allows quantitative detection of said antigen;
- e) said contacting is with a sample comprising said antibody; or
- f) said contacting allows quantitative detection of said antibody.

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11. A composition comprising:

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- a) a sterile binding compound of Claim 7, or
- b) said binding compound of Claim 7 and a carrier, wherein said carrier is:
 - i) an aqueous compound, including water, saline, and/or buffer; and/or
 - ii) formulated for oral, rectal, nasal, topical, or parenteral administration.

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12. An isolated or recombinant nucleic acid encoding said DCRS2 polypeptide of Claim 1, wherein said:

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- a) DCRS2 is from a human; or
- b) said nucleic acid:
 - i) encodes an antigenic peptide sequence of Table 1;
 - ii) encodes a plurality of antigenic peptide sequences of Table 1;
 - iii) exhibits identity over at least thirteen nucleotides to a natural cDNA encoding said segment;
 - iv) is an expression vector;
 - v) further comprises an origin of replication;
 - vi) is from a natural source;
 - vii) comprises a detectable label;
 - viii) comprises synthetic nucleotide sequence;
 - ix) is less than 6 kb, preferably less than 3 kb;

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- x) is from a primate;
- xi) comprises a natural full length coding sequence;
- xii) is a hybridization probe for a gene encoding said DCRS2; or
- xiii) is a PCR primer, PCR product, or mutagenesis primer.

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13. A cell or tissue comprising said recombinant nucleic acid of Claim 12.

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14. The cell of Claim 13, wherein said cell is:

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- a) a prokaryotic cell;
- b) a eukaryotic cell;
- c) a bacterial cell;
- d) a yeast cell;
- e) an insect cell;
- f) a mammalian cell;
- g) a mouse cell;
- h) a primate cell; or
- i) a human cell.

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15. A kit comprising said nucleic acid of Claim 12, and:

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- a) a compartment comprising said nucleic acid;
- b) a compartment further comprising a primate DCRS2 polypeptide; or
- c) instructions for use or disposal of reagents in said kit.

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16. A nucleic acid which:

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- a) hybridizes under wash conditions of 30 minutes at 30° C and less than 2M salt to the coding portion of SEQ ID NO: 1; or
- b) exhibits identity over a stretch of at least about 30 nucleotides to a primate DCRS2.

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17. The nucleic acid of Claim 16, wherein:

- a) said wash conditions are at 45° C and/or 500 mM salt; or

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b) said stretch is at least 55 nucleotides.

18. The nucleic acid of Claim 16, wherein:

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a) said wash conditions are at 55° C and/or 150 mM salt;
or

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b) said stretch is at least 75 nucleotides.

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19. A method of modulating physiology or development of a cell or tissue culture cells comprising contacting said cell with an agonist or antagonist of a mammalian DCRS2.

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20. The method of Claim 19, wherein said cell is transformed with a nucleic acid encoding a DCRS2 and another cytokine receptor subunit.

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	30	35	40	
cat ttt tat aaa aat ggc atc aaa gaa aga ttt caa atc aca agg att				240
His Phe Tyr Lys Asn Gly Ile Lys Glu Arg Phe Gln Ile Thr Arg Ile	45	50	55	
aat aaa aca aca gct cgg ctt tgg tat aaa aac ttt ctg gaa cca cat				288
Asn Lys Thr Thr Ala Arg Leu Trp Tyr Lys Asn Phe Leu Glu Pro His	60	65	70	
gct tct atg tac tgc act gct gaa tgt ccc aaa cat ttt caa gag aca				336
Ala Ser Met Tyr Cys Thr Ala Glu Cys Pro Lys His Phe Gln Glu Thr	75	80	85	
ctg ata tgt gga aaa gac att tct tct gga tat ccg cca gat att cct				384
Leu Ile Cys Gly Lys Asp Ile Ser Ser Gly Tyr Pro Pro Asp Ile Pro	90	95	100	105
gat gaa gta acc tgt gtc att tat gaa tat tca ggc aac atg act tgc				432
Asp Glu Val Thr Cys Val Ile Tyr Glu Tyr Ser Gly Asn Met Thr Cys	110	115	120	
acc tgg aat gct ggg aag ctc acc tac ata gac aca aaa tac gtg gta				480
Thr Trp Asn Ala Gly Lys Leu Thr Tyr Ile Asp Thr Lys Tyr Val Val	125	130	135	
cat gtg aag agt tta gag aca gaa gaa gag caa cag tat ctc acc tca				528
His Val Lys Ser Leu Glu Thr Glu Glu Glu Gln Gln Tyr Leu Thr Ser	140	145	150	
agc tat att aac atc tcc act gat tca tta caa ggc ggc aag aag tac				576
Ser Tyr Ile Asn Ile Ser Thr Asp Ser Leu Gln Gly Lys Lys Tyr	155	160	165	
ttg gtt tgg gtc caa gca gca aac gca cta ggc atg gaa gag tca aaa				624
Leu Val Trp Val Gln Ala Ala Asn Ala Leu Gly Met Glu Glu Ser Lys	170	175	180	185
caa ctg caa att cac ctg gat gat ata gtg ata cct tct gca gcc gtc				672
Gln Leu Gln Ile His Leu Asp Asp Ile Val Ile Pro Ser Ala Ala Val	190	195	200	
att tcc agg gct gag act ata aat gct aca gtg ccc aag acc ata att				720
Ile Ser Arg Ala Glu Thr Ile Asn Ala Thr Val Pro Lys Thr Ile Ile	205	210	215	
tat tgg gat agt caa aca aca att gaa aag gtt tcc tgt gaa atg aga				768
Tyr Trp Asp Ser Gln Thr Thr Ile Glu Lys Val Ser Cys Glu Met Arg	220	225	230	
tac aag gct aca aca aac caa act tgg aat gtt aaa gaa ttt gac acc				816
Tyr Lys Ala Thr Thr Asn Gln Thr Trp Asn Val Lys Glu Phe Asp Thr	235	240	245	
aat ttt aca tat gtg caa cag tca gaa ttc tac ttg gag cca aac att				864
Asn Phe Thr Tyr Val Gln Gln Ser Glu Phe Tyr Leu Glu Pro Asn Ile	250	255	260	265
aag tac gta ttt caa gtg aga tgt caa gaa aca ggc aaa agg tac tgg				912
Lys Tyr Val Phe Gln Val Arg Cys Gln Glu Thr Gly Lys Arg Tyr Trp	270	275	280	

cag cct tgg agt tca ccg ttt ttt cat aaa aca cct gaa aca gtt ccc 960
 Gln Pro Trp Ser Ser Pro Phe Phe His Lys Thr Pro Glu Thr Val Pro
 285 290 295

cag gtc aca tca aaa gca ttc caa cat gac aca tgg aat tct ggg cta 1008
 Gln Val Thr Ser Lys Ala Phe Gln His Asp Thr Trp Asn Ser Gly Leu
 300 305 310

aca gtt gct tcc atc tct aca ggg cac ctt act tct gac aac aga gga 1056
 Thr Val Ala Ser Ile Ser Thr Gly His Leu Thr Ser Asp Asn Arg Gly
 315 320 325

gac att gga ctt tta ttg gga atg atc gtc ttt gct gtt atg ttg tca 1104
 Asp Ile Gly Leu Leu Leu Gly Met Ile Val Phe Ala Val Met Leu Ser
 330 335 340 345

att ctt tct ttg att ggg ata ttt aac aga tca ttc ccg aac tgg gat 1152
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His Ile Trp Val Glu Pro Ala Thr Ile Phe Lys Met Gly Met Asn Ile
 10 15 20 25

Ser Ile Tyr Cys Gln Ala Ala Ile Lys Asn Cys Gln Pro Arg Lys Leu
 30 35 40

His Phe Tyr Lys Asn Gly Ile Lys Glu Arg Phe Gln Ile Thr Arg Ile
 45 50 55

Asn Lys Thr Thr Ala Arg Leu Trp Tyr Lys Asn Phe Leu Glu Pro His
 60 65 70

Ala Ser Met Tyr Cys Thr Ala Glu Cys Pro Lys His Phe Gln Glu Thr
 75 80 85

Leu Ile Cys Gly Lys Asp Ile Ser Ser Gly Tyr Pro Pro Asp Ile Pro
 90 95 100 105

Asp Glu Val Thr Cys Val Ile Tyr Glu Tyr Ser Gly Asn Met Thr Cys
 110 115 120

Thr Trp Asn Ala Gly Lys Leu Thr Tyr Ile Asp Thr Lys Tyr Val Val
 125 130 135

His Val Lys Ser Leu Glu Thr Glu Glu Glu Gln Gln Tyr Leu Thr Ser

aayatgacnt gyacntggaa ygcnggnaar ytnacntaya thgayacnaa rraygtngtn 480
 caygtnaarw snytngarac ngargarger carcartayy tnacnwsnws ntayathaay 540
 athwsnacng aywsnytnca rggnggnaar aartayyng tntgggtncn rgcngcnaay 600
 gcnytnggna tggargarws naarcarytn carathcayy tngaygayat hgtathccn 660
 wsnccngcng tnathwsnmq ngcngaracn athaaygca cngtncnaa racnathath 720
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 acnaaycara cntggaaygt naargartty gayacnaayt tyacntaygt ncarcarwsn 840
 garttytayy tngarccnaa yathaartay gtnttycarg tnmngtgyca rgaracnggn 900
 aarmntayt ggcrcctng gwsnwsnccn ttytycaya aracncnga racngtncn 960
 cargtnacnw snaargcmtt ycarcaygay acntggaayw snggnytnac ngtngcnwsn 1020
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 35 40 45
 Thr Leu Leu Ile Gly Ser Ser Leu Leu Ala Thr Cys Ser Val His Gly
 50 55 60
 Asp Pro Pro Gly Ala Thr Ala Glu Gly Leu Tyr Trp Thr Leu Asn Gly
 65 70 75 80
 Arg Arg Leu Pro Pro Glu Leu Ser Arg Val Leu Asn Ala Ser Thr Leu
 85 90 95
 Ala Leu Ala Leu Ala Asn Leu Asn Gly Ser Arg Gln Arg Ser Gly Asp
 100 105 110
 Asn Leu Val Cys His Ala Arg Asp Gly Ser Ile Leu Ala Gly Ser Cys
 115 120 125
 Leu Tyr Val Gly Leu Pro Pro Glu Lys Pro Val Asn Ile Ser Cys Trp
 130 135 140

6

Ser Lys Asn Met Lys Asp Leu Thr Cys Arg Trp Thr Pro Gly Ala His
 145 150 155 160
 Gly Glu Thr Phe Leu His Thr Asn Tyr Ser Leu Lys Tyr Lys Leu Arg
 165 170 175
 Trp Tyr Gly Gln Asp Asn Thr Cys Glu Glu Tyr His Thr Val Gly Pro
 180 185 190
 His Ser Cys His Ile Pro Lys Asp Leu Ala Leu Phe Thr Pro Tyr Glu
 195 200 205
 Ile Trp Val Glu Ala Thr Asn Arg Leu Gly Ser Ala Arg Ser Asp Val
 210 215 220
 Leu Thr Leu Asp Ile Leu Asp Val Val Thr Thr Asp Pro Pro Pro Asp
 225 230 235 240
 Val His Val Ser Arg Val Gly Gly Leu Glu Asp Gln Leu Ser Val Arg
 245 250 255
 Trp Val Ser Pro Pro Ala Leu Lys Asp Phe Leu Phe Gln Ala Lys Tyr
 260 265 270
 Gln Ile Arg Tyr Arg Val Glu Asp Ser Val Asp Trp Lys Val Val Asp
 275 280 285
 Asp Val Ser Asn Gln Thr Ser Cys Arg Leu Ala Gly Leu Lys Pro Gly
 290 295 300
 Thr Val Tyr Phe Val Gln Val Arg Cys Asn Pro Phe Gly Ile Tyr Gly
 305 310 315 320
 Ser Lys Lys Ala Gly Ile Trp Ser Glu Trp Ser His Pro Thr Ala Ala
 325 330 335
 Ser Thr Pro Arg Ser Glu Arg Pro Gly Pro Gly Gly Gly Ala Cys Glu
 340 345 350
 Pro Arg Gly Gly Glu Pro Ser Ser Gly Pro Val Arg Arg Glu Leu Lys
 355 360 365
 Gln Phe Leu Gly Trp Leu Lys Lys His Ala Tyr Cys Ser Asn Leu Ser
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 Thr Arg Asn Gln Val Leu Pro Asp Lys Leu
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 <213> rodent

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 Asp Pro Thr Leu Leu Ile Gly Ser Ser Leu Gln Ala Thr Cys Ser Ile
 35 40 45
 His Gly Asp Thr Pro Gly Ala Thr Ala Glu Gly Leu Tyr Trp Thr Leu
 50 55 60
 Asn Gly Arg Arg Leu Pro Ser Leu Ser Arg Leu Leu Asn Thr Ser Thr
 65 70 75 80
 Leu Ala Leu Ala Leu Ala Asn Leu Asn Gly Ser Arg Gln Gln Ser Gly
 85 90 95
 Asp Asn Leu Val Cys His Ala Arg Asp Gly Ser Ile Leu Ala Gly Ser
 100 105 110
 Cys Leu Tyr Val Gly Leu Pro Pro Glu Lys Pro Phe Asn Ile Ser Cys
 115 120 125
 Trp Ser Arg Asn Met Lys Asp Leu Thr Cys Arg Trp Thr Pro Gly Ala
 130 135 140
 His Gly Glu Thr Phe Leu His Thr Asn Tyr Ser Leu Lys Tyr Lys Leu
 145 150 155 160
 Arg Trp Tyr Gly Gln Asp Asn Thr Cys Glu Glu Tyr His Thr Val Gly
 165 170 175
 Pro His Ser Cys His Ile Pro Lys Asp Leu Ala Leu Phe Thr Pro Tyr
 180 185 190
 Glu Ile Trp Val Glu Ala Thr Asn Arg Leu Gly Ser Ala Arg Ser Asp
 195 200 205
 Val Leu Thr Leu Asp Val Leu Asp Val Val Thr Thr Asp Pro Pro Pro
 210 215 220
 Asp Val His Val Ser Arg Val Gly Gly Leu Glu Asp Gln Leu Ser Val
 225 230 235 240
 Arg Trp Val Ser Pro Pro Ala Leu Lys Asp Phe Leu Phe Gln Ala Lys
 245 250 255
 Tyr Gln Ile Arg Tyr Arg Val Glu Asp Ser Val Asp Trp Lys Val Val
 260 265 270
 Asp Asp Val Ser Asn Gln Thr Ser Cys Arg Leu Ala Gly Leu Lys Pro
 275 280 285
 Gly Thr Val Tyr Phe Val Gln Val Arg Cys Asn Pro Phe Gly Ile Tyr
 290 295 300
 Gly Ser Lys Lys Ala Gly Ile Trp Ser Glu Trp Ser His Pro Thr Ala
 305 310 315 320
 Ala Ser Thr Pro Arg Ser Glu Arg Pro Gly Pro Gly Gly Gly Val Cys
 325 330 335
 Glu Pro Arg Gly Gly Glu Pro Ser Ser Gly Pro Val Arg Arg Glu Leu

210 215 220
 Ser Ser Phe Phe Ile Arg Asp Ile Ile Lys Pro Asp Pro Pro Lys Asn
 225 230 235 240
 Leu Gln Leu Lys Pro Leu Lys Asn Ser Arg Gln Val Glu Val Ser Trp
 245 250 255
 Glu Tyr Pro Asp Thr Trp Ser Thr Pro His Ser Tyr Phe Ser Leu Thr
 260 265 270
 Phe Cys Val Gln Val Gln Gly Lys Ser Lys Arg Glu Lys Lys Asp Arg
 275 280 285
 Val Phe Thr Asp Lys Thr Ser Ala Thr Val Ile Cys Arg Lys Asn Ala
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 305 310 315 320
 Glu Trp Ala Ser Val Pro Cys Ser
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 <213> rodent

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 35 40 45
 Thr Cys Asp Thr Pro Glu Glu Asp Asp Ile Thr Trp Thr Ser Asp Gln
 50 55 60
 Arg His Gly Val Ile Gly Ser Gly Lys Thr Leu Thr Ile Thr Val Lys
 65 70 75 80
 Glu Phe Leu Asp Ala Gly Gln Tyr Thr Cys His Lys Gly Gly Glu Thr
 85 90 95
 Leu Ser His Ser His Leu Leu Leu His Lys Lys Glu Asn Gly Ile Trp
 100 105 110
 Ser Thr Glu Ile Leu Lys Asn Phe Lys Asn Lys Thr Phe Leu Lys Cys
 115 120 125
 Glu Ala Pro Asn Tyr Ser Gly Arg Phe Thr Cys Ser Trp Leu Val Gln
 130 135 140
 Arg Asn Met Asp Leu Lys Phe Asn Ile Lys Ser Ser Ser Ser Ser Pro
 145 150 155 160
 Asp Ser Arg Ala Val Thr Cys Gly Met Ala Ser Leu Ser Ala Glu Lys

10

165 170 175

Val Thr Leu Asp Gln Arg Asp Tyr Glu Lys Tyr Ser Val Ser Cys Gln
 180 185 190

Glu Asp Val Thr Cys Pro Thr Ala Glu Glu Thr Leu Pro Ile Glu Leu
 195 200 205

Ala Leu Glu Ala Arg Gln Gln Asn Lys Tyr Glu Asn Tyr Ser Thr Ser
 210 215 220

Phe Phe Ile Arg Asp Ile Ile Lys Pro Asp Pro Pro Lys Asn Leu Gln
 225 230 235 240

Met Lys Pro Leu Lys Asn Ser Gln Val Glu Val Ser Trp Glu Tyr Pro
 245 250 255

Asp Ser Trp Ser Thr Pro His Ser Tyr Phe Ser Leu Lys Phe Phe Val
 260 265 270

Arg Ile Gln Arg Lys Lys Glu Lys Met Lys Glu Thr Glu Glu Gly Cys
 275 280 285

Asn Gln Lys Gly Ala Phe Leu Val Glu Lys Thr Ser Thr Glu Val Gln
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Cys Lys Gly Gly Asn Val Cys Val Gln Ala Gln Asp Arg Tyr Tyr Asn
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<400> 8

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Gln Cys His Ala Ser Arg Tyr Pro Val Ala Val Asp Cys Ser Trp Thr
 35 40 45

Pro Leu Gln Ala Pro Asn Ser Thr Arg Ser Thr Ser Phe Ile Ala Thr
 50 55 60

Tyr Arg Leu Gly Val Ala Thr Gln Gln Gln Ser Gln Pro Cys Leu Gln
 65 70 75 80

Arg Ser Pro Gln Ala Ser Arg Cys Thr Ile Pro Asp Val His Leu Phe
 85 90 95

Ser Thr Val Pro Tyr Met Leu Asn Val Thr Ala Val His Pro Gly Gly
 100 105 110

Ala Ser Ser Ser Leu Leu Ala Phe Val Ala Glu Arg Ile Ile Lys Pro

12

165

170

175

Phe His Arg Val Gly Pro Ile Glu Ala Thr Ser Phe Ile Leu Arg Ala
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 Met Ser Leu Gly Lys
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 Pro Gly Val Ser Ala Gly Thr Pro Val Ser Trp Phe Arg Asp Gly Asp
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 Ser Arg Leu Leu Gln Gly Pro Asp Ser Gly Leu Gly His Arg Leu Val
 65 70 75 80
 Leu Ala Gln Val Asp Ser Pro Asp Glu Gly Thr Tyr Val Cys Gln Thr
 85 90 95
 Leu Asp Gly Val Ser Gly Gly Met Val Thr Leu Lys Leu Gly Phe Pro
 100 105 110
 Pro Ala Arg Pro Glu Val Ser Cys Gln Ala Val Asp Tyr Glu Asn Phe
 115 120 125
 Ser Cys Thr Trp Ser Pro Gly Gln Val Ser Gly Leu Pro Thr Arg Tyr
 130 135 140
 Leu Thr Ser Tyr Arg Lys Lys Thr Leu Pro Gly Ala Glu Ser Gln Arg
 145 150 155 160
 Glu Ser Pro Ser Thr Gly Pro Trp Pro Cys Pro Gln Asp Pro Leu Glu
 165 170 175
 Ala Ser Arg Cys Val Val His Gly Ala Glu Phe Trp Ser Glu Tyr Arg
 180 185 190
 Ile Asn Val Thr Glu Val Asn Ser Leu Gly Ala Ser Thr Cys Leu Leu
 195 200 205
 Asp Val Arg Leu Gln Ser Ile Leu Arg Pro Asp Pro Pro Gln Gly Leu

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Leu	Asp	Gly	Ala	Leu	Gly	Gly	Thr	Val	Thr	Leu	Gln	Leu	Gly	Tyr	Pro				
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Arg	Val	Glu	Ser	Val	Pro	Gly	Tyr	Pro	Arg	Arg	Leu	Arg	Ala	Ser	Trp				
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Thr	Tyr	Pro	Ala	Ser	Trp	Pro	Cys	Gln	Pro	His	Phe	Leu	Leu	Lys	Phe				
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Arg	Leu	Gln	Tyr	Arg	Pro	Ala	Gln	His	Pro	Ala	Trp	Ser	Thr	Val	Glu				
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Pro	Ala	Gly	Leu	Glu	Glu	Val	Ile	Thr	Asp	Ala	Val	Ala	Gly	Leu	Pro				
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His	Ala	Val	Arg	Val	Ser	Ala	Arg	Asp	Phe	Leu	Asp	Ala	Gly	Thr	Trp				
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Glu	Pro	Gln	Val	Asp	Ser	Pro	Ala	Pro	Pro	Arg	Pro	Ser	Leu	Gln	Pro				
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His	Pro	Arg	Leu	Leu	Asp	His	Arg	Asp	Ser	Val	Glu	Gln	Val	Ala	Val				
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Leu	Ala	Ser	Leu	Gly	Ile	Leu	Ser	Phe	Leu	Gly	Leu	Val	Ala	Gly	Ala				
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Leu Ala Leu Gly Leu Trp Leu Arg Leu Arg Arg Gly Gly Lys Asp Gly
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Pro Gly Ala Pro Asn Leu
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Gly Val Glu Pro Glu Asp Asn Ala Thr Val His Trp Val Leu Arg Lys
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Pro Ala Ala Gly Ser His Pro Ser Arg Trp Ala Gly Met Gly Arg Arg
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Tyr Arg Ala Gly Arg Pro Ala Gly Thr Val His Leu Leu Val Asp Val
 100 105 110

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Asn Val Val Cys Glu Trp Gly Pro Arg Ser Thr Pro Ser Leu Thr Thr
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Lys Ala Val Leu Leu Val Arg Lys Phe Gln Asn Ser Pro Ala Glu Asp
 145 150 155 160

Phe Gln Glu Pro Cys Gln Tyr Ser Gln Glu Ser Gln Lys Phe Ser Cys
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Gln Leu Ala Val Pro Gln Gly Asp Ser Ser Phe Tyr Ile Val Ser Met
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Cys Val Ala Ser Ser Val Gly Ser Lys Phe Ser Lys Thr Gln Thr Phe
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Gln Gly Cys Gly Ile Leu Gln Pro Asp Pro Pro Ala Asn Ile Thr Val
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Thr Ala Val Ala Arg Asn Pro Arg Trp Leu Ser Val Thr Trp Gln Asp
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 Pro Lys Leu Ser Cys Phe Arg Lys Asn Pro Leu Val Asn Ala Ile Cys
 115 120 125
 Glu Trp Arg Pro Ser Ser Thr Pro Ser Pro Thr Thr Lys Ala Val Leu
 130 135 140
 Phe Ala Lys Lys Ile Asn Thr Thr Asn Gly Lys Ser Asp Phe Gln Val
 145 150 155 160
 Pro Cys Gln Tyr Ser Gln Gln Leu Lys Ser Phe Ser Cys Gln Val Glu
 165 170 175
 Ile Leu Glu Gly Asp Lys Val Tyr His Ile Val Ser Leu Cys Val Ala
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 Asn Ser Val Gly Ser Lys Ser Ser His Asn Glu Ala Phe His Ser Leu
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 Lys Met Val Gln Pro Asp Pro Pro Ala Asn Leu Val Val Ser Ala Ile
 210 215 220
 Pro Gly Arg Pro Arg Trp Leu Lys Val Ser Trp Gln His Pro Glu Thr
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 Val Trp Ser Lys Glu Phe Thr Val Leu Leu Leu Pro Val Ala Gln Tyr
 260 265 270
 Gln Cys Val Ile His Asp Ala Leu Arg Gly Val Lys His Val Val Gln
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 Pro Glu Val Thr Gly Thr Pro Trp Ile Ala Glu Pro Arg Thr Thr Pro
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 Ala Gly Ile Leu Trp Asn Pro Thr Gln Val Ser Val Glu Asp Ser Ala
 325 330 335
 Asn His Glu Asp Gln Tyr Glu Ser Ser Thr Glu Ala Thr Ser Val Leu
 340 345 350
 Ala Pro Val Gln Glu Ser Ser Ser Met Ser Leu Pro Thr Phe Leu Val
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Ala Gly Gly Ser Leu Ala Phe Gly Leu Leu Leu Cys Val Phe Ile Ile
 370 375 380

Leu Arg Leu Lys Gln Lys Trp Lys Ser Glu Ala Glu Lys Glu Ser Lys
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Thr Thr Ser Pro Pro Pro Pro Pro Tyr Ser Leu Gly Pro Leu Lys Pro
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Thr Phe Leu Leu Val Pro Leu Leu Thr Pro His Ser Ser Gly Ser Asp
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Asn Thr Val Asn His Ser Cys Leu Gly Val Arg Asp Ala Gln Ser Pro
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Tyr Asp Asn Ser Asn Arg Asp Tyr Leu Phe Pro Arg
 450 455 460

INTERNATIONAL SEARCH REPORT

National Application No

PCT/US 00/14867

A. CLASSIFICATION OF SUBJECT MATTER		
IPC 7 C12N15/12 C07K14/715 C07K16/28 A61K38/17		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed by classification symbols)		
IPC 7 C12N C07K		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the International search (name of data base and, where practical, search terms used)		
STRAND, MEDLINE, BIOSIS, EPO-Internal		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE SWISSPROT 'Online! Sequence P40190: Interleukin-6 receptor beta chain precursor, membrane glycoprotein GP130 from rat; 1 February 1995 (1995-02-01) XP002152341 compare residues 122-129 with residues 120-127 in SEQ ID NO:2, and residues 201-204 with residues 200-203 in SEQ ID NO:2	1-3,12, 16-18
A	WO 99 20755 A (ELSON G ET AL; GLAXO GROUP LIMITED) 29 April 1999 (1999-04-29) the mGP130 sequence in Figure 2	1-20
A	US 5 716 804 A (MOORE KW ET AL. SCHERING CORPORATION) 10 February 1998 (1998-02-10) the whole document	1-20
<input type="checkbox"/> Further documents are listed in the continuation of box C. <input checked="" type="checkbox"/> Patent family members are listed in annex.		
* Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "S" document member of the same patent family		
Date of the actual completion of the international search		Date of mailing of the international search report
9 November 2000		22/11/2000
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016		Authorized officer Cupido, M

INTERNATIONAL SEARCH REPORT
Information on patent family members

International Application No
PCT/US 00/14867

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9920755 A	29-04-1999	AU 1334799 A EP 1027438 A	10-05-1999 16-08-2000
US 5716804 A	10-02-1998	NONE	